

Antimicrobial and anticancer potential of soil bacterial metabolites - a comprehensive and updated review

A. Ram Kumar^{1,2*}, S. Kumaresan²

¹Department of Plant Biology and Plant Biotechnology, Thiruthangal Nadar College, Selavayal, Chennai- 600051, India.

²PG and Research Department of Plant Biology and Plant Biotechnology, Ramakrishna Mission Vivekananda College, Mylapore, Chennai- 600004, India.

ARTICLE INFO

Article history:

Received on: March 26, 2023

Accepted on: June 27, 2023

Available online: August 10, 2023

Key words:

Bacteria,
Bioactivity,
Natural products,
Soil microbes,
Pseudomonas spp.,
Streptomyces spp.

ABSTRACT

The majority of natural products currently used in the medical field are derived from microbial or plant sources. The bioactive compounds derived from natural sources exhibit tremendous structural and chemical diversity. According to previous research, only a small percentage of the world's plant and microbial diversity has been examined for bioactivities. The compounds originating from secondary metabolites of microorganisms are more useful for the development of novel drugs due to their biological friendliness and drug-likeness than any other compounds. Thus, recent research suggests that microorganisms obtained from diverse habitats and natural resources offer various bioactive secondary metabolites with incredibly wider chemical entities, hopefully, an alternative remedy for many diseases. Soil bacteria are capable of producing a variety of natural bioactive compounds for the treatment of various diseases. The three genera *Bacillus* spp., *Streptomyces* spp., and *Pseudomonas* spp. have been the prime focus to produce different types of antibiotics. However, to date, there are no reviews that evaluated the antimicrobial and anticancer properties of soil bacterial metabolites. Hence, the current review aimed to assess the antimicrobial and anticancer potential of soil bacterial metabolites.

1. INTRODUCTION

Man is dependent on natural products to maintain good health and protect against various diseases from time immemorial. Natural products are the richest source for drug discovery and currently, 65% of the approved drugs in medical fields are obtained from them [1,2]. The database of natural products contains more than 210,000 biologically active compounds with abundant chemical diversity [3]. In the year 2013, 1453 new compounds had been approved by the United States Food and Drug Administration of which approximately 40% were derived from natural products and their derivatives [4]. Scientific communities have given more importance to natural products as drugs derived from them provide better treatment compared to synthetic products [5]. Besides, the compounds derived from natural sources contain abundant structural diversity compared to synthetic compounds and play an important role in new drug discoveries [6]. Especially, compounds derived from microorganisms play a significant role in treating infectious diseases and cancer [2,7]. Among microorganisms, bacteria and fungi are the main candidates focused on the production of bioactive compounds [8,9], as they

have existed on earth for billions of years and have evolved many biosynthetic pathways by novel mechanisms to synthesize secondary metabolites. The various applications of microbial natural products are demonstrated in Figure 1. The discovery of bioactive compounds from microbes involves various steps, including isolation, structural elucidation, and establishing the biosynthetic pathway leading to the formation of secondary metabolites [7,9].

According to an estimate, 0.1% of bacterial species and 5% of fungal species of the world are known to man, which only a small fraction has been screened for bioactivity [10,11]. Antimicrobial agents isolated from actinomycetes include streptomycin, gentamycin and rifamycin, whereas anti-cancer agents comprise mitomycin, aclarubicin, neocarzinostatin, doxorubicin, mithramycin, and carzinophilin [12]. Previously, there are no reviews that evaluated the antimicrobial and anticancer properties of soil bacterial metabolites. Therefore, the current review aimed to assess the antimicrobial and anticancer potential of soil bacterial metabolites.

2. CANCER-A DEADLY DISEASE

In recent years, the incidence of cancer is increasing at a phenomenal rate. According to a World Health Organization (WHO) estimate, cancer affects approximately 10 million people by 2020, or nearly one in six deaths. Breast, lung, colon, rectum, and prostate cancers are the most frequent cancers. Tobacco use, a high body mass index, alcohol consumption, and a lack of physical activity account for almost one-third of cancer fatalities. In low- and lower-middle-income

*Corresponding Author:

Dr. A. Ram Kumar

Department of Plant Biology and Plant Biotechnology,

Thiruthangal Nadar College,

Selavayal, Chennai- 600 004, India.

Email: ramramkumar185@gmail.com

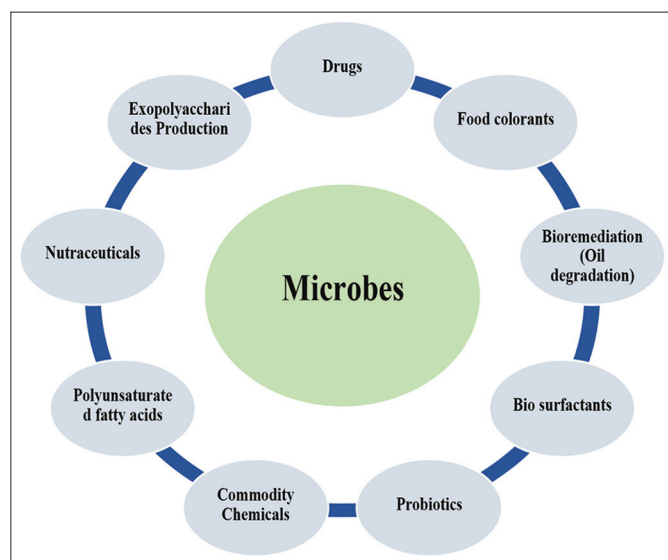


Figure 1: Various applications of microbial natural products.

countries, cancer-causing diseases such as the human papillomavirus and hepatitis account for roughly 30% of cancer cases. Many tumors are curable if identified early and treated properly [13,14].

Based on the statistical report of global cancer, approximately 12.7 million cases of cancer were detected and 7.6 million cancer mortality occurred in the year 2011 [15,16] and 8.8 million of death in 2015 [17]. Cancer is caused by both external (chemicals, radiation, tobacco, alcohol abuse, and infectious organisms) and internal (familial history, hormones, immune conditions, inherited mutations, and mutation occur from metabolism) factors. The other factors that cause cancer include infections 15–20%, diet, obesity 30–35%, tobacco 25–30%, stress, lack of physical activity, radiation, and environmental pollutants [18].

3. ANTIMICROBIAL RESISTANCE (AMR)

Antimicrobial agents have saved millions of lives from microbial infections for more than seven decades. However, AMR is a major problem faced by the medical world and this has caused adverse effects on human beings [19]. Pathogens are regularly becoming resistant towards antimicrobial agents. This problem is further increased due to the independent and continuous use of antibiotics that cause serious health diseases in humans [20]. Recently, the AMR reports predicted that due to multidrug-resistant (MDR) infections the death rate will be 10 million in the year 2050 [21]. It referred to the evolution of pathogenic microbes such as fungi, bacteria and viruses that developed resistance against the antimicrobial drugs [22,23]. At present, 60% of the Gram-negative bacteria have developed resistance to all classes of antimicrobial agents including carbapenems, cephalosporin, and fluoroquinolones. [24,25]. Similarly, Gram-positive bacteria *Staphylococcus aureus* significantly increased its resistance to the antibiotic methicillin and referred to as methicillin-resistant *S. aureus* [26]. According to the world health organization report in 2017, three groups of bacterial genus namely *Pseudomonas*, *Acinetobacter*, and various *Enterobacteriaceae* members including *Escherichia coli*, *Klebsiella*, *Proteus*, and *Serratia* developed resistance to most of the antibiotics [27-29].

The fungal infection is yet another serious issue in the medical field and has the potential to harm every individual. Approximately, 1.2 billion people are suffering from various fungal diseases universally [30]. The

most common disease-causing pathogenic fungi are *Aspergillus* and *Candida*. The infection caused by *Aspergillus* in human is extremely alarming. Particularly, *Aspergillus niger* causes various diseases to humans which includes pulmonary diseases, fungal ear infections, temporary hearing loss, and skin diseases [31]. The *Candida* spp. is the common cause of fungal nosocomial bloodstream infections. Among the *Candida* spp. *Candida albicans* is the most prevalent species involved in nosocomial bloodstream infections, gastrointestinal tract infection, mucosal oral cavity diseases, intestinal infections, and skin problems [32].

4. NEED FOR ALTERNATIVE DRUGS TO TREAT CANCER AND MICROBIAL INFECTIONS

The available cancer therapies such as radiation therapy, surgery, chemotherapy, hormone therapy, and immunotherapy often fail to achieve complete cancer remission. These types of treatments cause significant side effects to humans, such as blood clots, hair loss, pain, anemia, fatigue, thrombocytopenia, diarrhea, constipation, neurological complications, and unpleasant to fatal infections [33,34]. Moreover, the currently available anticancer drugs are limited in their safety and efficacy. Successful cancer therapy depends on its preferential wipeout of cancer cells without side effects or negligible toxicity to the normal cells [35,36].

After the revolution, in the 1960s the “golden era,” most of the important antibiotics such as cephalosporins, macrolides, tetracyclines, and aminoglycosides were discovered and major problems of chemotherapy have been solved. The history is being repeated now because the exciting compounds are losing their efficacy due to the increase in AMR and failure to treat the MDR bacteria and fungi. It becomes a universal problem for public health [37,38]. In addition, the widely used antifungal drugs like azoles cause significant side effects and frequently decrease the efficacy against fungal pathogens [39]. For this reason, the discovery of new bioactive compounds with safety and efficacy to treat cancer and microbial infections is an exclusively important objective.

5. SOIL MICROORGANISMS

Soil is a rich reservoir of microorganisms that differentially favors the diversity of microbes according to the geographical region and environmental factors. The soil microbes are the key components of the forest biomes and they are playing a vital role in the soil aggregation, nitrogen fixation and cycling of nutrients through lignin and cellulose breakdown [40,41]. Soil microbes, such as fungi and bacteria control the ecosystem by decomposition and maintain the health of the ecosystem. The majority of the currently used drugs are derived from the soil microorganisms belonging to the genera *Bacillus*, *Streptomyces*, *Micromonospora*, *Penicillium*, and *Cephalosporium* [42]. More than 500 bioactive compounds are discovered every year among which almost 60% is obtained from the soil microbes [43,44].

6. SOIL BACTERIA

The soil bacteria are the key contributors to productivity in the ecosystem and nutrient cycling. It is the most diverse and abundant microbial community in soil [45]. Besides, soil bacteria are the main source of production of bioactive secondary metabolites with enormous biological properties [46,47]. It has been used for a variety of applications ranging from crop production to drug discovery. Members of the genus *Bacillus* are commonly found in soil and produce a variety

of bioactive secondary metabolites that are effective against a variety of life-threatening diseases [42]. As per literature, many bacterial strains isolated from soil samples, for example, *Bacillus pumillus*, *Bacillus lentus*, *Enterobacter aerogenes*, *Bacillus alvei*, *Micrococcus roseus*, and *Bacillus amyloliquefaciens* have shown strong bioactivity against the various pathogens [48-50]. Nowadays, most of the research workers focused on soil microbial communities and diversity of soil bacteria especially from *Bacillus* communities [51]. The isolation of pure compounds from bacterial strains is not a simple process. Because each crude extract contains several compounds, constructing a crude extracts library takes less time than constructing a purified compounds library, and the range of diversity may be comparable to that of a large systematic compound library. To detect the target component in the various compound mixtures, however, a sophisticated and robust screening technology with high specificity and sensitivity is required. To eliminate known and redundant compounds, suggest which peak should be efficiently purified, and accumulate knowledge for structure determination, a dereplication technique is necessary [52]. The dereplication process for bacterial natural product screening method is shown in Figure 2.

7. BACTERIAL METABOLITES

Bacterial secondary metabolites are a major source of the drugs that lead, with attractive bioactivities. In particular, a variety of secondary metabolites is produced by bacteria, generating approximately half of the discovered antibiotics [53]. In the group of bacteria, the *Streptomyces*, *Pseudomonas*, and *Bacillus* species are the most frequent producer of bioactive compounds. Among these, *Bacillus* strains have a great potential to use in various application fields including antibiotics, enzymes producers, vitamins, probiotics, and bioprotection products. They also contributed significant roles in the biodegradation of pollutants in the environment [54].

The most prominent species of *Bacillus* including *Bacillus licheniformis*, *Bacillus subtilis*, *Bacillus circulans*, *Bacillus amyloliquefaciens*, *Bacillus polymixa*, *Bacillus pumilus*, and *Bacillus cereus* [55,56].

These organisms commonly produce isocoumarins, lipopeptides, polyketides, aminoglycoside, aminopolyol, phospholipids, phosphonoligopeptide, and terpenoids [57,58]. Similarly, the most important antibiotics rifamycin and gramicidin are reported from *Streptomyces* and *Bacillus* [59]. American physician William Coley reported that the bacterial strains had produced several safe vaccines against carcinomas, lymphomas, sarcomas, and melanomas cancers [60]. This has prompted the development of many new drugs from bacteria for various diseases [61]. The process involved in the isolated of bioactive compounds is shown in Figure 3.

8. ANTIMICROBIAL METABOLITES FROM SOIL BACTERIA

Today both industrialists and academicians focus interest on soil bacteria due to its discrete advantages over the other microbes. The soil bacteria not only produce unique bioactive metabolites but also commercially important natural products [62]. They are employed in various fields such as fermentation process (cheese, brewing, baking and butter manufacturing), chemical manufacturing (acetone, ethanol, organic acid, perfume, enzymes, etc.), and drug discovery. Therefore, the application of soil bacteria's open up new areas of biotechnological exploitations, which lead to the essentials of isolation and cultivation of these organisms [63,64]. They produce a variety of vaccines, antibiotics, steroids, and therapeutically useful compounds with strong biological activities [65]. Streptomycin was the first aminoglycoside antibiotic derived from the soil bacterium *Streptomyces griseus*. Following that, important antibiotics such as macrolide, chloramphenicol, glycopeptide (e.g., vancomycin), and tetracycline were derived from the soil bacteria [66,67].

Bacillus is the most common bacteria found in the soil and many species of this genus produce a variety of antimicrobial compounds [68]. The bactericidal and fungicidal compounds derived from the soil *Bacillus* spp. includes megacin from *Bacillus megaterium* [69], polyfermentacin from *Bacillus polyfermenticus* [70],

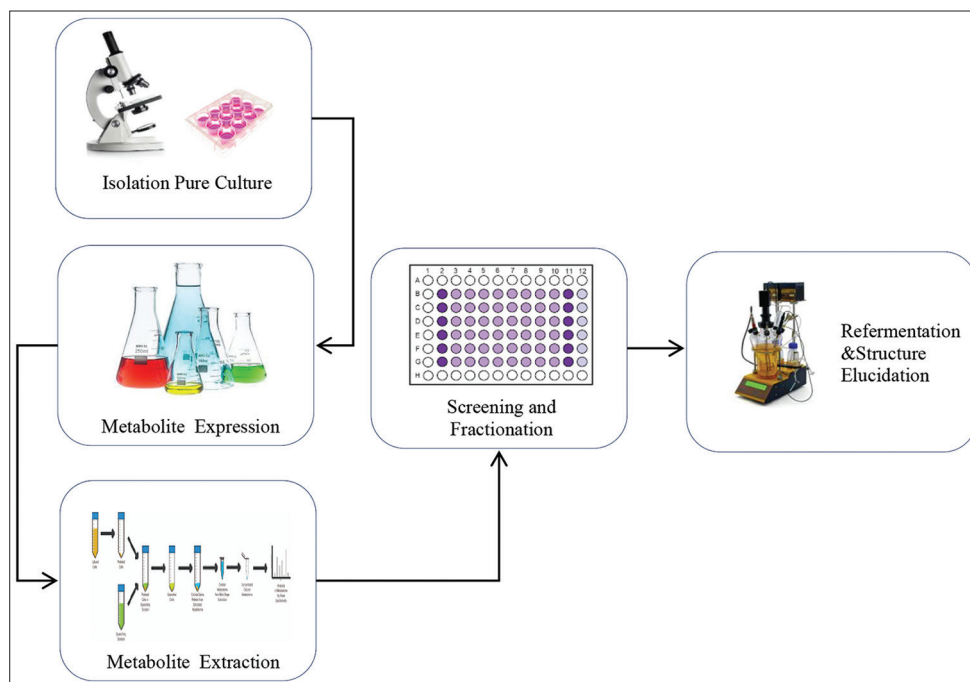


Figure 2: Dereplication process for bacterial natural product screening.

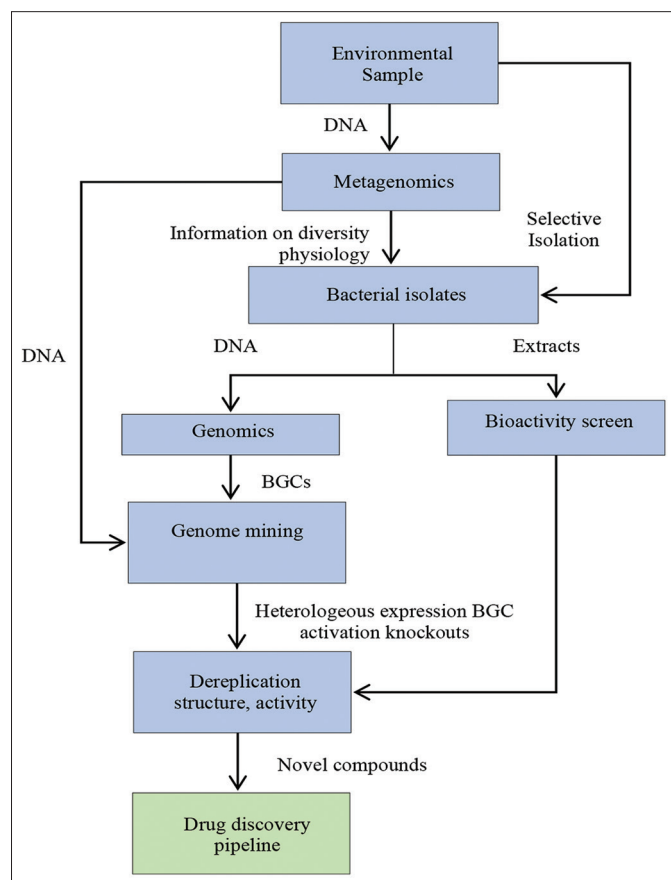


Figure 3: Overview of the workflow for the discovery of bioactive compounds from bacteria.

cerin and zwittermixin 14 from *B. cereus* [71], bacilysin 1, subtilin, ericin, mersacidin, sublancin, subtilolysin, amicoumacin 4 and iturin 7 produced by *B. subtilis* [72], lichenin from *Bacillus licheniformis*, diffidin 10, bacilysin 1, macrolactin 12, iturin 7 from *B. amyloliquefaciens* [73], amicoumacin 4 and bacilysin 1 from *B. pumilus* [74], Zwittermixin 14, thuricin, tochicin, kurstakin, and entomocin from *Bacillus thuringiensis* [71]. In addition to these, mitomycin C an antitumor compound derived from *Streptomyces caespitosus* was discovered by Japanese microbiologists in the year 1950. The mitomycin C is used in the treatment of lungs, breast, hepatic carcinoma, head, neck, bladder, and colorectal cancers [75].

Erythromycin isolated from the soil bacterium *Saccharopolyspora erythraea* is the first generation of macrolide and the macrolide antibiotics potential against both Gram-positive and Gram-negative bacteria [76]. The compound cyclopentapeptides derived from *Streptomyces flavovirens* isolated from the Antarctic soil samples showed potent antitumor and antimicrobial activity [77,78] reported laterosporulin 10 (LS10) a type of anticancer agent derived from *Brevibacillus* spp. showed anticancer activity against five different human cell lines such as lung carcinoma (H1299), cervical cancer (HeLa), breast cancer (MCF-7), fibrosarcoma (HT1080), and embryonic kidney cancer (HEK293T). Similarly, 700 antibiotics have been derived from *Micromonospora* spp., including 150 aminoglycosides [79]. Hover *et al.*, [80] reported a new class of antibiotic malacidins discovered from the soil bacteria, which are active against different types of MDR pathogens. The list of antimicrobial compounds isolated from bacterial strains is detailed in Table 1.

Singh and Wahla [127] disclosed the secondary metabolites extracted from *Streptomyces werraensis* KBR01 isolated from the rhizosphere soil samples showed antifungal activity against *Fusarium oxysporum*. Recently, it was found by the disc diffusion method that several *Streptomyces* spp. isolated from the rhizosphere soil showed excellent antifungal activity against *C. albicans* [128]. Likewise, Andargie and Li, [129] revealed antifungal compounds extracted from *Streptoverticillium morookaense* isolated from the soil samples of pine forest at China, showed potential antimicrobial activity against the tested pathogens *Ustilaginoidea virens*, *Bipolaris maydis* and *Rhizoctonia solani*. Adlin Jenifer *et al.*, [130] reported the strain *Nocardioopsis* sp. isolated from the soil samples collected from Kovalamsolar salterns, Tamil Nadu, showed antimicrobial activity against *E. coli*, *Pseudomonas aeruginosa*, *A. niger*, etc. Pattnaik [131] described that the strain *Micromonospermae chinospora* isolated from the soil samples of Western Odisha, showed potential antibacterial activity against *S. aureus*. A study by Mahdiyah *et al.* [132] isolated several bacterial strains from peat soil samples showed strong antibacterial activity against methicillin-susceptible *S. aureus* (ATCC 29213) and *E. coli* (ATCC 35218). Kumar *et al.* [1] isolated the bacterial strain *B. subtilis* from soil samples were collected from Avalanche reserve forest of south zone Nilgiris district. The bioactive metabolites of the strain showed strong antimicrobial activity against *E. aerogenes*, *Enterococcus faecalis*, *Alcaligenes faecalis* and *C. albicans*. Zhou *et al.* [133] isolated several *Bacillus* strains from rhizosphere soil that have antimicrobial activity against *Phytophthora infestans*, *R. solani*, *Pseudomonas syringae*, *Erwinia carotovora*, *Verticillium dahlia* and *Botrytis cinerea*. Recently, a study by Osama *et al.* [12] isolated the four *Streptomyces* spp. (SH8, SH10, SH12, and SH13) from the top layer of agricultural soil in Beni-Suef Governorate, Egypt. All the four isolated showed a broad spectrum of antimicrobial activity against *C. albicans* (ATCC 60193), *Listeria monocytogenes* (ATCC 7644), *E. coli* (clinical isolate), *Salmonella enterica* (ATCC 14028), *B. subtilis* (environmental sample), and *S. aureus* (ATCC 43300).

9. ANTICANCER METABOLITES FROM SOIL BACTERIA

The anticancer metabolites isolated from the soil bacterial strains can be considered as a safe alternative for synthetic drugs. Malkov *et al.* [134] reported a Gram-positive bacteria *B. oligonitrophilus* KU-1 isolated from the soil samples of Kazan city, Russia exhibited the effective anticancer activity against colon cancer cell line. Abraham *et al.* [135] isolated antitumor compound doxorubicin from the soil bacterium *Streptomyces peucetius* var. caesius which showed anticancer activity against skin cancer. Similarly, a new bacterial strain AAA5 isolated from the humus soil of Western Ghats, India, has been identified as *Streptomyces aurantiacus*. It produced a quinone-related antibiotic resistomycin showing potential anticancer activity against HeLa (cervical carcinoma) and HepG2 (hepatic carcinoma) cell lines with growth inhibition 0.005 and 0.006 g/mL, respectively [136].

Rebecamycin is a *Lentzea aerocolonigenes*-derived indolocarbazole-based antitumor antibiotic. Rebecamycin was discovered in a Panama soil sample from an actinomycete culture of C-38383. Different study groups called it *Saccharothrix aerocolonigenes* and *Lechevalieria aerocolonigenes* after it was first placed under *Streptomyces* sp. and titled *Nocardia aerocolonigenes* [137]. Hou *et al.* [138] reported *Streptomyces* sp. isolated from the soil samples collected at Huiquan Square in China, produced a new polyketide glycoside, gilvocarcin HE showing moderate anticancer activity against P388, K562 and MCF-7 cell lines with IC_{50} values of 45, 39, and 36 μ g/mL, respectively. Similarly, Balachandran *et al.* [139] reported the bacteria ERI-135

Table 1: List of antimicrobial compounds isolated from bacterial strains.

Bacteria	Compound	Biological effects	References
<i>Bacillus megaterium</i>	Megacin	Bactericidal	[81]
<i>Bacillus cereus</i>	Cerein	Bactericidal	[82]
<i>Bacillus subtilis</i> , <i>Bacillus pumilus</i>	Amicoumacin 4	Antibacterial	[83]
<i>Bacillus amyloliquefaciens</i>	Bacillaene 11	Antibacterial	[84]
<i>Pseudomonas fluorescens</i>	Nunapeptin, Nunamycin	Antifungal	[85]
<i>Scaphirhynchus albus</i>	Malacidin	Antimicrobial	[80]
<i>Escherichia coli</i>	Cadaside	Antibacterial	[86]
<i>Streptomyces malaysiensis</i>	Azalomycin F	Antifungal	[87]
<i>Streptomyces</i> spp. MA37	Accramycin A	Antibacterial	[88]
<i>Amycolatopsis</i> spp. MST-108494	Amycolatopsins A–C	Antibacterial	[89]
<i>Bacillus polyfermenticus</i>	Polyfermenticin	Antimicrobial	[90]
<i>Bacillus</i> spp.	Bogorol A	Antibacterial	[91]
<i>Bacillus</i> spp.	Loloatin B	Antibacterial	[92]
<i>Bacillus amyloliquefaciens</i>	Macrolactin S	Antibacterial	[93]
<i>Bacillus amyloliquefaciens</i>	Macrolactin V	Antibacterial	[93]
<i>Bacillus laterosporus</i>	Basiliskamides	Antifungal	[94]
<i>Streptomyces misionensis</i>	Streptenols	Antibacterial	[95]
<i>Streptomyces</i> spp.	Dibohemamines	Antimicrobial	[96]
<i>Thermoactinomyces vulgaris</i>	Thermoactinoamide A	Antimicrobial	[97]
<i>Streptomyces clavuligerus</i>	Cephalosporins	Antibacterial	[98]
<i>Streptomyces cattleya</i>	Thienamycin	Antibacterial	[99]
<i>Streptomyces erythraea</i>	Erythromycin	Antibacterial	[100]
<i>Streptomyces orientalis</i>	Vancomycin	Antibacterial	[101]
<i>Streptomyces griseus</i>	Streptomycin	Antibacterial	[102]
<i>Marantochloa purpurea</i>	Gentamycin	Antibacterial	[103]
<i>Streptomyces</i> spp.,	Tetracyclines	Antibacterial	[104]
<i>Pseudomonas fluorescens</i>	Mupirocin	Antibacterial	[105]
<i>Streptomyces roseosporus</i>	Daptomycin	Antibacterial	[106]
<i>Streptomyces nodosus</i>	AmphotericinB	Antifungal	[107]
<i>Nocardiosis alba</i>	Z)-1-((1-hydroxypenta-2,4-dien1-yl) oxy) anthracene-9,10-dione	Antibacterial	[108]
<i>Streptomyces</i> spp. HW-003	AMRSA1	Antibacterial	[109]
<i>Streptomyces</i> spp.C34	Chaxamycins	Antibacterial	[110]
<i>Streptomyces kanamyceticus</i>	Bekanamycin	Antibacterial	[111]
<i>Streptomyces triticiradicis</i> sp.	2,3-dihydroxybutanone	Antifungal	[112]
<i>Streptomyces fradiae</i>	Neomycin	Antibacterial	[113]
<i>Streptomyces venezuelae</i>	Chloramphenicol	Antibacterial	[114]
<i>Streptomyces griseus</i>	Albomycin	Antibacterial	[115]
<i>Streptomyces laurentii</i>	Thiostrepton	Antibacterial	[116]
<i>Streptomyces</i> spp.	Clindamycin	Antibacterial	[117]
<i>Actinoplanes</i> spp. ATCC33706	Ramoplanin	Antibacterial	[118]
<i>Verrucosipora</i> AB-18-032	Abyssomicins	Antibacterial	[119]
<i>Micromonospora</i> spp.	Micromonosporin	Antibacterial	[120]
<i>Nocardia</i> spp.	Thiolactomycin	Antibacterial	[121]
<i>Streptomyces aureofaciens</i>	Chlortetracycline	Antibacterial	[122]
<i>Streptomyces rimosus</i>	Oxytetracycline	Antibacterial	[123]
<i>Streptomyces mediodicicus</i>	Mediomycin B	Antifungal	[124]
<i>Kibdelosporangium aridum</i>	Aridicins A, BandC	Antimicrobial	[125]
<i>Streptomyces</i> spp.	Bonactin	Antibacterial	[126]

Table 2: List of anticancer compounds isolated from bacterial strains.

Bacteria	Compound	Biological effects	References
<i>Kibdelosporangium</i> spp.	Kibdelon	Anticancer	[146]
<i>Bacillus</i> spp. CND-914	Halobacillin	Anticancer	[147]
<i>Bacillus</i> spp.	Mixirin	Anticancer	[148]
<i>Bacillus silvestris</i>	Bacillistatins	Anticancer	[149]
<i>Streptomyces verticillus</i>	Bleomycin	Anticancer	[150]
<i>Microbacterium mangrovin</i> MUSC115	(3R,8aS)-3-methyl-1,2,3,4,6,7,8,8a-octahydropyrrolo[1,2-a] pyrazine-1,4-dione	Anticancer	[151]
<i>Streptomyces peuceitius</i>	Doxorubicin	Anticancer	[152]
<i>Streptomyces vinaceus</i>	Citreamicindelta	Anticancer	[153]
<i>Micromonospora chersina</i>	Dynemicin	Anticancer	[154]
<i>Streptomyces peuceitius</i>	Amrubicin	Anticancer	[155]
<i>Micromonospermae chinospora</i> spp.	Gemtuzumabozogamicin	Anticancer	[156]
<i>Streptomyces</i> spp. KML-2	Chromomycinand 1-(1H-indol-3-yl)-propane-1,2,3-triol	Anticancer	[157]
<i>Streptomyces</i> spp. VN1	Furan-type	Anticancer	[158]
<i>Streptomyces bulli</i> GJA1	Angucyclinones	Anticancer	[159]
<i>Streptomyces aurantiacus</i> AAA5	Resistomycin	Anticancer	[136]
<i>Streptomyces peuceitius</i> and <i>Streptomyces</i> spp.	Daunorubicin	Anticancer	[160]

identified as *Methylobacterium* spp. isolated from the Doddabetta forest soil showed anticancer activity against human lung cancer cell line (A 549). The antitumor compound glycopeptides mixture of antibiotics bleomycin isolated from *Streptomyces verticillus* has been used in the treatment of ovarian cancer, head carcinomas, testicular carcinomas, and neck carcinomas [140,141].

Parthiban *et al.* [142] reported *B. thuringiensis* S13 isolated from the soil of Mandapam, coastal area South India, produced exopolymer with strong anticancer activity against lung cancer cell line (A549) with an IC₅₀ value of 133.27 µg/mL. Kumar *et al.* [143] reported the medicinal valuable unknown compounds were derived from the two soil bacterial strains *B. pumilus* and *B. cereus* by a fractionated method. All the isolated fractions were tested against liver cancer cell lines by MTT assay, showed high anticancer activity. Xu *et al.* [144] reported the natural compound 7-Cyano-7-deazaguanine isolated from the soil bacterium *Streptomyces qinglanensis* showed potent antitumor activity against HepG2 and HeLa cell lines. Similarly, another anticancer compound bovocin HC5 derived from the bacterium *Streptomyces bovis* divulged cytotoxicity against human liver hepatocellular carcinoma (HepG2) and human breast cancer (MCF7) [145]. The list of anticancer compounds isolated from bacterial strains is detailed in Table 2.

The biosurfactant produced by *P. aeruginosa* isolated from the oil-contaminated soil showed anticancer activity against HeLa cell lines by significantly controlling cell proliferation of the cells [161]. The compound 5-methyl phenazine-1-carboxylic acid betaine, derived from the *P. putida* soil bacterium, exhibited promising anticancer activity against human breast cancer and lung cancer cell lines [162]. In addition, Ramasubburayan *et al.* [163] reported that crude extract of *B. subtilis* RG isolated from the soil samples of the Southeast coast of India showed significant antitumor activity against human breast cancer cell lines (MCF7). Kim *et al.* [164] reported that the metabolites of bacterial strain *B. amyloliquefaciens* isolated from the rhizosphere soil of Korean ginseng showed strong antiproliferative and anticancer activity against the colorectal cancer cell lines of

humans such as HT-29, LoVo, SW480, and HCT116. Recently, a study by Osama *et al.* [12] isolated the four *Streptomyces* spp. (SH8, SH10, SH12, and SH13) from the top layer of agricultural soil in Beni-Suef Governorate, Egypt. Among the four isolates, the isolates SH4 and SH12 showed anti-cancer activity against breast cells MCF-10A and the hepatoma cell line hepatoma G2 (HepG2). In addition, Kumar *et al.* [165] reported that the bioactive metabolites of soil bacterium *B. subtilis* showed significant anticancer activity against breast cancer cell line (MCF-7) by MTT assay. The *in silico* analysis showed the compound metaraminol having the maximum docking score -7.27 Kcal/mol against the breast cancer targeted protein estrogen receptor alpha (ER α).

10. CONCLUSION AND FUTURE DIRECTION

The current review highlights the antimicrobial and anticancer properties of soil bacterial metabolites demonstrated by existing studies. For scientists, finding potent secondary metabolite producers like soil bacteria is an exciting and demanding platform. Members of soil bacteria generate industrially valuable compounds such as enzymes, antibiotics, and pigments despite living under extreme circumstances. In its early phases, the use of soil microorganisms as a hotspot for novel bioactive optional metabolites. Therefore, more, research is required for the identification of novel soil bacterial strains as well as novel compounds from them.

Around the world, new potent small molecules with significant anticancer potential and a manageable safety profile are desperately needed. Many anticancer medications now being used in clinical trials have a variety of side effects. Reduced toxicity in non-targeted tissues and an improved site-targeted strategy are urgently needed. As a result, there is a significant need for bioactive cytotoxic natural compounds, which are preferable to produced molecules. Blockbuster anticancer drugs such as camptothecin, doxorubicin, topotecan, vinblastine, saviincristine, paclitaxel, and others have been used in nano-based platforms such as polymeric nanoparticles, polymer-drug conjugates, dendrimers, liposomes, and immunoliposomes to

improve targeted tissue delivery while reducing toxicity to healthy cells. Nature's small molecules, as well as synthetic chemicals, have made major contributions to today's commercially available pharmaceuticals. Natural compounds have a minor advantage over manufactured products due to their lower toxicity profile, despite their large contribution. It's vital to remember that drugs based on natural compounds are not completely free of side effects. Actinomycin D's approval for the treatment of particular cancers has been delayed due to its side effects, which include tissue necrosis, myelosuppression, dermatotoxicity, and gastrointestinal enterotoxicity. Computational approaches for discovering new natural compounds and chemical derivatization give a broad platform for developing anticancer drugs in the near future. In view of all of these aspects, natural products hold a lot of promise for the development of novel drug seeds to address unmet needs in cancer therapy.

11. AUTHORS' CONTRIBUTIONS

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agreed to be accountable for all aspects of the work. All the authors are eligible to be an author as per the International Committee of Medical Journal Editors (ICMJE) requirements/guidelines.

12. FUNDING

There is no funding to report.

13. CONFLICTS OF INTEREST

The authors report no financial or any other conflicts of interest in this work.

14. ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

15. DATA AVAILABILITY

All data generated and analyzed are included within this review article.

16. PUBLISHER'S NOTE

This journal remains neutral with regard to jurisdictional claims in published institutional affiliation.

REFERENCES

- Ram Kumar A, Ramesh R, Thamilvanan D, Stephen A, Kumaresan S. *In vitro* antimicrobial activity, optimization of bioactive secondary metabolites and molecular characterization of *B. subtilis* isolated from soils of Western Ghats, India. *Int Res J Pharm* 2019;10:205-12.
- Atanasov AG, Zotchev SB, Dirsch VM, International Natural Product Sciences Taskforce, Supuran CT. Natural products in drug discovery: Advances and opportunities. *Nat Rev Drug Discov* 2021;20:200-16.
- Rodrigues T, Reker D, Schneider P, Schneider G. Counting on natural products for drug design. *Nat Chem* 2016;8:531-41.
- Patridge E, Gareiss P, Kinch MS, Hoyer D. An analysis of FDA-approved drugs: Natural products and their derivatives. *Drug Discov Today* 2016;21:204-7.
- Rahman M, Sarker SD. *Antimicrobial Natural Products*. Netherlands: Elsevier; 2020. p. 77-113. Available from: <https://www.linkinghub.elsevier.com/retrieve/pii/S0065774320300270> [Last accessed on 2022].
- Balachandra C, Padhi D, Govindaraju T. Cyclic dipeptide: A privileged molecular scaffold to derive structural diversity and functional utility. *ChemMedChem* 2021;16:2558-87.
- Karthikeyan A, Joseph A, Nair BG. Promising bioactive compounds from the marine environment and their potential effects on various diseases. *J Genet Eng Biotechnol* 2022;20:14.
- Mishra S, Goyal D, Phurailatpam L. Targeted 16S rRNA gene and ITS2 amplicon sequencing of leaf and spike tissues of *Piper longum* identifies new candidates for bioprospecting of bioactive compounds. *Arch Microbiol* 2021;203:3851-67.
- Tran PN, Yen MR, Chiang CY, Lin HC, Chen PY. Detecting and prioritizing biosynthetic gene clusters for bioactive compounds in bacteria and fungi. *Appl Microbiol Biotechnol* 2019;103:3277-87.
- Jenssen M. *Bioprospecting of Marine Microorganisms for the Discovery of Antibacterial Compounds-Isolation, Structure Elucidation and Bioactivity Assessment of Marine Microbial Natural Products*. Norway: UiT The Arctic University of Norway; 2022.
- Thomas AT, Rao JV, Subrahmanyam VM, Chandrashekar HR, Maliyakkal N, Kisan TK, *et al.* *In vitro* anticancer activity of microbial isolates from diverse habitats. *Braz J Pharm Sci* 2011;47:279-87.
- Osama N, Bakeer W, Raslan M, Soliman HA, Abdelmohsen UR, Sebak M. Anti-cancer and antimicrobial potential of five soil *Streptomyces*: A metabolomics-based study. *R Soc Open Sci* 2022;9:211509.
- World Health Organization. *Cancer Fact Sheet*. Geneva: World Health Organization; 2022. Available from: <https://www.who.int/news-room/fact-sheets/detail/cancer> [Last accessed on 2022].
- Muthaiyan L, Arockia Jayalatha JBB, Noel N. An Introduction to Cancer Biomarkers. *Biomarkers and Biosensors for Cervical Cancer Diagnosis* 2021.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011;61:69-90.
- Siegel RL, Miller KD, Jemal A. *Cancer statistics, 2018*. *CA Cancer J Clin* 2018;68:7-30.
- World Health Organization. *Media Centre*. Geneva: World Health Organization; 2018.
- Dukre MT, Shinde JS, Bhavar AR. A review: Comprehensive study on anticancer therapy. *Int J Res Trends Innov* 2021;2021:6101-9.
- Wallinga D, Smit LA, Davis MF, Casey JA, Nachman KE. A review of the effectiveness of current US policies on antimicrobial use in meat and poultry production. *Curr Environ Health Rep* 2022;9:339-54.
- Uddin TM, Chakraborty AJ, Khusro A, Zidan BR, Mitra S, Bin ET, *et al.* Antibiotic resistance in microbes: History, mechanisms, therapeutic strategies and future prospects. *J Infect Public Health* 2021;14:1750-66.
- Ramírez-Castillo FY, Moreno-Flores AC, Avelar-González FJ, Márquez-Díaz F, Harel J, Guerrero-Barrera AL. An evaluation of multidrug-resistant *Escherichia coli* isolates in urinary tract infections from Aguascalientes, Mexico: Cross-sectional study. *Ann Clin Microbiol Antimicrob* 2018;17:34.
- Mihankhah A, Khoshbakht R, Raeisi M, Raeisi V. Prevalence and antibiotic resistance pattern of bacteria isolated from urinary tract infections in Northern Iran. *J Res Med Sci* 2017;22:108.
- Tenney J, Hudson N, Alnifaify H, Li JT, Fung KH. Risk factors for acquiring multidrug-resistant organisms in urinary tract infections: A systematic literature review. *Saudi Pharm J* 2018;26:678-84.
- Hawkey PM, Warren RE, Livermore DM, McNulty CA, Enoch DA, Otter JA, *et al.* Treatment of infections caused by multidrug-resistant Gram-negative bacteria: Report of the British Society for Antimicrobial Chemotherapy/Healthcare Infection Society/British

- Infection Association Joint Working Party. *J Antimicrob Chemother* 2018;73:iii2-78.
25. Rajivgandhi G, Maruthupandy M, Ramachandran G, Priyanga M, Manoharan N. Detection of ESBL genes from ciprofloxacin resistant Gram negative bacteria isolated from urinary tract infections (UTIs). *Front Lab Med* 2018;2:5-13.
 26. Nepal K, Pant ND, Neupane B, Belbase A, Baidhya R, Shrestha RK, *et al.* Extended spectrum beta-lactamase and metallo beta-lactamase production among *Escherichia coli* and *Klebsiella pneumoniae* isolated from different clinical samples in a tertiary care hospital in Kathmandu, Nepal. *Ann Clin Microbiol Antimicrob* 2017;16:62.
 27. Sands K, Carvalho MJ, Portal E, Thomson K, Dyer C, Akpulu C, *et al.* Characterization of antimicrobial-resistant Gram-negative bacteria that cause neonatal sepsis in seven low-and middle-income countries. *Nat Microbiol* 2021;6:512-23.
 28. Shaikh S, Fatima J, Shakil S, Rizvi SM, Kamal MA. Antibiotic resistance and extended spectrum beta-lactamases: Types, epidemiology and treatment. *Saudi J Biol Sci* 2015;22:90-101.
 29. Kumar M, Curtis A, Hoskins C. Application of nanoparticle technologies in the combat against anti-microbial resistance. *Pharmaceutics* 2018;10:11.
 30. Ibe C. The fight against mycoses in Africa: Are we making progress? *Clin Microbiol Infect* 2022;28:9-12.
 31. Merad Y, Derrar H, Belmokhtar Z, Belkacemi M. *Aspergillus* genus and its various human superficial and cutaneous features. *Pathogens* 2021;10:643.
 32. Talapko J, Juzbašić M, Matijević T, Pustijanac E, Bekić S, Kotris I, *et al.* *Candida albicans*-the virulence factors and clinical manifestations of infection. *J Fungi* 2021;7:79.
 33. Aggarwal S, Verma SS, Aggarwal S, Gupta SC. Drug repurposing for breast cancer therapy: Old weapon for new battle. *Semin Cancer Biol* 2021;68:8-20.
 34. Mansouri V, Beheshtizadeh N, Gharibshahian M, Sabouri L, Varzandeh M, Rezaei N. Recent advances in regenerative medicine strategies for cancer treatment. *Biomed Pharmacother* 2021;141:111875.
 35. Samal P, Begum S. Drug loaded nanomaterials for hematological malignancies diagnosis and enhanced targeted therapy. In: *Advanced Nanomaterials for Point of Care Diagnosis and Therapy*. Netherlands: Elsevier; 2022. p. 383-98. Available from: <https://www.linkinghub.elsevier.com/retrieve/pii/B9780323857253000167> [Last accessed on 2022].
 36. Haider K, Yar MS. *Advances of Benzimidazole Derivatives as Anticancer Agents: Bench to Bedside*. London, U.K: Intechopen; 2022. Available from: <https://www.intechopen.com/online-first/79835> [Last accessed on 2022].
 37. Jangra M, Kaur P, Tambat R, Raka V, Mahey N, Chandal N, *et al.* Recent Updates on Bacterial Secondary Metabolites to Overcome Antibiotic Resistance in Gram-Negative Superbugs: Encouragement or Discontinuation? *Antimicrobial Resistance*. Singapore: Springer Singapore; 2022. p. 385-418. Available from: https://www.link.springer.com/10.1007/978-981-16-3120-7_14 [Last accessed on 2022].
 38. Fisher MC, Alastruey-Izquierdo A, Berman J, Bicanic T, Bignell EM, Bowyer P, *et al.* Tackling the emerging threat of antifungal resistance to human health. *Nat Rev Microbiol* 2022;20:557-71.
 39. Balcerak MI, Stewart AG, Chapman P, Lazarus S. Reducing the off-target endocrinologic adverse effects of azole antifungals-can it be done? *Int J Antimicrob Agents* 2022;59:106587.
 40. Wang C, Masoudi A, Wang M, Yang J, Yu Z, Liu J. Land-use types shape soil microbial compositions under rapid urbanization in the Xiong'an New Area, China. *Sci Total Environ* 2021;777:145976.
 41. Zhang J, Li J, Fan Y, Mo Q, Li Y, Li Y, *et al.* Effect of nitrogen and phosphorus addition on litter decomposition and nutrients release in a tropical forest. *Plant Soil* 2020;454:139-53.
 42. Cobongela SZ. *Applications of Microbes in Antibiotics*. Singapore: Springer; 2022. p. 693-710. Available from: https://www.link.springer.com/10.1007/978-981-16-2225-0_25 [Last accessed on 2022].
 43. Molinari G. Natural products in drug discovery: Present status and perspectives. *Adv Exp Med Biol* 2009;655:13-27.
 44. Kang MK, Kim JH, Liu MJ, Jin CZ, Park DJ, Kim J, *et al.* New discovery on the nematode activity of aureothin and alloaureothin isolated from endophytic bacteria *Streptomyces* sp. AE170020. *Sci Rep* 2022;12:3947.
 45. Jiao S, Xu Y, Zhang J, Hao X, Lu Y. Core microbiota in agricultural soils and their potential associations with nutrient cycling. *ASM J* 2019;4:30.
 46. Al-Shaibani MM, Mohamed RM, Sidik NM, El Enshasy HA, Al-Gheethi A, Noman E, *et al.* Biodiversity of secondary metabolites compounds isolated from phylum actinobacteria and its therapeutic applications. *Molecules* 2021;26:4504.
 47. Selim MS, Abdelhamid SA, Mohamed SS. Secondary metabolites and biodiversity of actinomycetes. *J Genet Eng Biotechnol* 2021;19:72.
 48. Olasinbo OB, Sylvanus CU, Peters OO. Antibiotic-producing bacteria isolated from some natural habitats in the Federal Capital Territory (FCT), Nigeria. *Afr J Microbiol Res* 2022;16:43-55.
 49. Abdulkadir M, Waliyu S. Screening and isolation of the soil bacteria for ability to produce antibiotics. *Eur J Appl Sci* 2012;4:211-5.
 50. Boottanun P, Potisap C, Hurdle JG, Sermswan RW. Secondary metabolites from *Bacillus amyloliquefaciens* isolated from soil can kill *Burkholderia pseudomallei*. *AMB Express* 2017;7:16.
 51. Bogati K, Walczak M. The impact of drought stress on soil microbial community, enzyme activities and plants. *Agronomy* 2022;12:189.
 52. Da H, Gan M, Jiang D, Xing C, Zhang Z, Fei L, *et al.* Epitaxial regeneration of spent graphite anode material by an eco-friendly in-depth purification route. *ACS Sustain Chem Eng* 2021;9:16192-202.
 53. Shalabi A, Eskander D, Badawe M. Isolation of secondary metabolites from marine *Streptomyces sparsus* ASD203 and evaluation its bioactivity. *Egypt J Chem* 2021;65:539-47.
 54. Jung S, Woo C, Fugaban JI, Bucheli JE, Holzapfel WH, Todorov SD. Bacteriocinogenic potential of *Bacillus amyloliquefaciens* isolated from Kimchi, a traditional Korean fermented cabbage. *Probiotics Antimicrob Proteins* 2021;13:1195-212.
 55. Kamali M, Guo D, Naeimi S, Ahmadi J. Perception of biocontrol potential of *Bacillus inaquosorum* KR2-7 against tomato fusarium wilt through merging genome mining with chemical analysis. *Biology (Basel)* 2022;11:137.
 56. Jeong SJ, Ryu MS, Yang HJ, Wu XH, Jeong DY, Park SM. Bacterial distribution, biogenic amine contents, and functionalities of traditionally made doenjang, a long-term fermented soybean food, from different Areas of Korea. *Microorganisms* 2021;9:1348.
 57. Qadir A, Hussain MM, Bin ZM, Hameed MA, Farooqi ZU. Unveiling the potential of *Bacillus* sp. in Bioremediation and Biocontrol. *Cham: Springer*; 2022. p. 519-38. Available from: https://www.link.springer.com/10.1007/978-3-030-85465-2_22 [Last accessed on 2022].
 58. Surovy MZ, Rahman S, Dame ZT, Islam T. Discovery of bioactive natural products from *Bacillus* species: Chemistry, Biosynthesis and Biological Activities. *Cham: Springer*; 2022. p. 47-87. Available from: https://www.link.springer.com/10.1007/978-3-030-85465-2_3 [Last accessed on 2022].
 59. Khabthani S, Rolain JM, Merhej V. *In silico/in vitro* strategies leading to the discovery of new nonribosomal peptide and polyketide antibiotics active against human pathogens. *Microorganisms* 2021;9:2297.
 60. Kucerova P, Cervinkova M. Spontaneous regression of tumour and

- the role of microbial infection-possibilities for cancer treatment. *Anticancer Drugs* 2016;27:269-77.
61. Terreni M, Taccani M, Pregnolato M. New antibiotics for multidrug-resistant bacterial strains: Latest research developments and future perspectives. *Molecules* 2021;26:2671.
 62. Findlay BL. The chemical ecology of predatory soil bacteria. *ACS Chem Biol* 2016;11:1502-10.
 63. Gayathiri G, Kiruthiga P, Karthikeyan R, Anand AV, Sivamurugan V, Saradhadevi KM. Enzymatic production of organic acids via microbial fermentative processes. In: *Biomass, Biofuels, Biochem*. Netherlands: Elsevier; 2022. p. 37-54.
 64. Ranghar S, Agrawal S, Agrawal PK. *Microbial Products: Protein, Enzyme, Secondary Metabolites and Chemicals*. Microbial Interview Agriculture Environment. Singapore: Springer Singapore; 2019. p. 347-84.
 65. Chitra J, Rajendren S, Jeyakanthan J, Samy BG, Mercy JJ, Manikandan N, *et al*. Microbes and their Products as Novel Therapeutics in Medical Applications. *Bioprospecting Microb Divers*. Netherlands: Elsevier; 2022. p. 203-21. Available from: <https://www.linkinghub.elsevier.com/retrieve/pii/B9780323909587000194> [Last accessed on 2022].
 66. Murugaiyan J, Kumar PA, Rao GS, Iskandar K, Hawser S, Hays JP, *et al*. Progress in alternative strategies to combat antimicrobial resistance: Focus on antibiotics. *Antibiotics* 2022;11:200.
 67. Mohr KL. History of antibiotics research. *Curr Top Microbiol Immunol* 2016;398:237-72.
 68. Chaudhary P, Sharma A, Chaudhary A, Khati P, Gangola S, Maithani D. Illumina based high throughput analysis of microbial diversity of maize rhizosphere treated with nanocompounds and *Bacillus* sp. *Appl Soil Ecol* 2021;159:103836.
 69. Basi-Chipalu S, Sthapit P, Dhital S. A review on characterization, applications and structure-activity relationships of *Bacillus* species-produced bacteriocins. *Drug Discov Ther* 2022;16:55-62.
 70. Lee NK, Kim WS, Paik HD. *Bacillus* strains as human probiotics: Characterization, safety, microbiome, and probiotic carrier. *Food Sci Biotechnol* 2019;28:1297-305.
 71. Ortiz A, Sansinenea E. Chemical compounds produced by *Bacillus* sp. Factories and their role in nature. *Mini Rev Med Chem* 2019;19:373-80.
 72. Engelbrecht G, Horak I, van Rensburg PJ, Claassens S. *Bacillus*-based bionematicides: Development, modes of action and commercialisation. *Biocontrol Sci Technol* 2018;28:629-53.
 73. Lv J, Da R, Cheng Y, Tuo X, Wei J, Jiang K, *et al*. Mechanism of antibacterial activity of *Bacillus amyloliquefaciens* C-1 Lipopeptide toward anaerobic *Clostridium difficile*. *Biomed Res Int* 2020;2020:3104613.
 74. Baranova MN, Kudzhaev AM, Mokrushina YA, Babenko VV, Kornienko MA, Malakhova MV, *et al*. Deep functional profiling of wild animal microbiomes reveals probiotic *Bacillus pumilus* strains with a common biosynthetic fingerprint. *Int J Mol Sci* 2022;23:1168.
 75. Bradner WT. Mitomycin C: A clinical update. *Cancer Treat Rev* 2001;27:35-50.
 76. Lü J, Long Q, Zhao Z, Chen L, He W, Hong J, *et al*. Engineering the erythromycin-producing strain *Saccharopolyspora erythraea* HOE107 for the heterologous production of polyketide antibiotics. *Front Microbiol* 2020;11:593217.
 77. Bratchkova A, Ivanova V. Bioactive metabolites produced by microorganisms collected in Antarctica and the Arctic. *Biotechnol Bioinform Equip* 2011;25:1-7.
 78. Baindara P, Gautam A, Raghava GP, Korpole S. Anticancer properties of a defensin like class IId bacteriocin Laterosporulin10. *Sci Rep* 2017;7:46541.
 79. Chen L, Wang Z, Du S, Wang G. Antimicrobial activity and functional genes of actinobacteria from Coastal Wetland. *Curr Microbiol* 2021;78:3058-67.
 80. Hover BM, Kim SH, Katz M, Charlop-Powers Z, Owen JG, Ternei MA, *et al*. Culture-independent discovery of the malacidins as calcium-dependent antibiotics with activity against multidrug-resistant Gram-positive pathogens. *Nat Microbiol* 2018;3:415-22.
 81. Lisboa MP, Bonatto D, Bizani D, Henriques JA, Brandelli A. Characterization of a bacteriocin-like substance produced by *Bacillus amyloliquefaciens* isolated from the Brazilian Atlantic forest. *Int Microbiol* 2006;9:111-8.
 82. Bizani D, Dominguez AP, Brandelli A. Purification and partial chemical characterization of the antimicrobial peptide cerein 8A. *Lett Appl Microbiol* 2005;41:269-73.
 83. Pinchuk IV, Bressollier P, Sorokulova IB, Verneuil B, Urdaci MC. Amicoumacin antibiotic production and genetic diversity of *B. subtilis* strains isolated from different habitats. *Res Microbiol* 2002;153:269-76.
 84. Chen XH, Koumoutsis A, Scholz R, Borriss R. More than anticipated-production of antibiotics and other secondary metabolites by *Bacillus amyloliquefaciens* FZB42. *J Mol Microbiol Biotechnol* 2009;16:14-24.
 85. Michelsen CF, Watrous J, Glaring MA, Kersten R, Koyama N, Dorrestein PC, *et al*. nonribosomal peptides, key biocontrol components for *Pseudomonas fluorescens* In5, isolated from a greenlandic suppressive soil. *mBio* 2015;6:e00079.
 86. Wu C, Shang Z, Lemetre C, Ternei MA, Brady SF, Cadasides, Calcium-dependent acidic lipopeptides from the soil metagenome that are active against multidrug-resistant bacteria. *J Am Chem Soc* 2019;141:3910-9.
 87. Cheng J. Azalomycin F complex is an antifungal substance produced by *Streptomyces malaysiensis* MJM1968 isolated from agricultural soil. *J Korean Soc Appl Biol Chem* 2010;53:545-52.
 88. Maglangit F, Fang Q, Leman V, Soldatou S, Ebel R, Kyeremeh K, *et al*. Accramycin A, a new aromatic polyketide, from the soil bacterium, *Streptomyces* sp. MA37. *Molecules* 2019;24:3384.
 89. Khalil ZG, Salim AA, Vuong D, Crombie A, Lacey E, Blumenthal A, *et al*. Amycolatopsins A-C: Antimycobacterial glycosylated polyketide macrolides from the Australian soil *Amycolatopsis* sp. MST-108494. *J Antibiot (Tokyo)* 2017;70:1097-103.
 90. Lee KH, Jun KD, Kim WS, Paik HD. Partial characterization of polyfermenticin SCD, a newly identified bacteriocin of *Bacillus polyfermenticus*. *Lett Appl Microbiol* 2001;32:146-51.
 91. Bao S, Wang H, Zhang W, Xie Z, Fang T. An investigation into the effects of silver nanoparticles on natural microbial communities in two freshwater sediments. *Environ Pollut* 2016;219:696-704.
 92. Yang Y, Quensen J, Mathieu J, Wang Q, Wang J, Li M, *et al*. Pyrosequencing reveals higher impact of silver nanoparticles than Ag⁺ on the microbial community structure of activated sludge. *Water Res* 2014;48:317-25.
 93. Abdu N, Abdullahi AA, Abdulkadir A. Heavy metals and soil microbes. *Environ Chem Lett* 2017;15:65-84.
 94. Ngoua-Meye-Misso RL, Sima-Obiang C, Ndong JD, Ondo JP, Abessolo F, Obame-Engonga LC. Phytochemical screening, antioxidant, anti-inflammatory and antiangiogenic activities of *Lophira procera* A. Chev. (*Ochnaceae*) medicinal plant from Gabon. *Egypt J Basic Appl Sci* 2018;5:80-6.
 95. Tarazona G, Schleissner C, Rodríguez P, Pérez M, Cañedo LM, Cuevas C. Streptenols F-I Isolated from the marine-derived *Streptomyces misionensis* BAT-10-03-023. *J Nat Prod* 2017;80:1034-8.
 96. Jiang B, Zhao W, Li S, Liu H, Yu L, Zhang Y, *et al*. Cytotoxic dibohemamines D-F from a *Streptomyces* species. *J Nat Prod* 2017;80:2825-9.
 97. Teta R, Marteinsson VT, Longeon A, Klonowski AM, Groben R,

- Bourguet-Kondracki ML, *et al.* Thermoactinoamide A, an antibiotic lipophilic cyclopeptide from the icelandic *Thermophilic bacterium Thermoactinomyces vulgaris*. J Nat Prod 2017;80:2530-5.
98. Aharonowitz Y, Demain AL. Carbon catabolite regulation of cephalosporin production in *Streptomyces clavuligerus*. Antimicrob Agents Chemother 1978;14:159-64.
 99. Sanada M, Miyano T, Iwadare S, Williamson JM, Arison BH, Smith JL, *et al.* Biosynthesis of fluorothreonine and fluoroacetic acid by the thienamycin producer, *Streptomyces cattleya*. J Antibiot (Tokyo) 1986;39:259-65.
 100. Chng C, Lum AM, Vroom JA, Kao CM. A key developmental regulator controls the synthesis of the antibiotic erythromycin in *Saccharopolyspora erythraea*. Proc Natl Acad Sci 2008;105:11346-51.
 101. Griffith RS. Vancomycin use--an historical review. J Antimicrob Chemother 1984;14:1-5.
 102. Waksman SA, Reilly HC, Johnstone DB. Isolation of streptomycin-producing strains of *Streptomyces griseus*. J Bacteriol 1946;52:393-7.
 103. Chu J, Li B, Zhang S, Li Y. On-line ultrasound stimulates the secretion and production of gentamicin by *Micromonospora echinospora*. Process Biochem 2000;35:569-72.
 104. Asagbra AE, Sanni AI, Oyewole OB. Solid-state fermentation production of tetracycline by *Streptomyces* strains using some agricultural wastes as substrate. World J Microbiol Biotechnol 2005;21:107-14.
 105. El-Sayed AK, Hothersall J, Cooper SM, Stephens E, Simpson TJ, Thomas CM. Characterization of the mupirocin biosynthesis gene cluster from *Pseudomonas fluorescens* NCIMB 10586. Chem Biol 2003;10:419-30.
 106. Penn J, Li X, Whiting A, Latif M, Gibson T, Silva CJ, *et al.* Heterologous production of daptomycin in *Streptomyces lividans*. J Ind Microbiol Biotechnol 2006;33:121-8.
 107. Abu-Salah KM. Amphotericin B: An update. Br J Biomed Sci 1996;53:122-33.
 108. Janardhan A, Kumar AP, Viswanath B, Saigopal DV, Narasimha G. Production of bioactive compounds by actinomycetes and their antioxidant properties. Biotechnol Res Int 2014;2014:217030.
 109. Kemung HM, Tan LT, Khan TM, Chan KG, Pusparajah P, Goh BH, *et al.* *Streptomyces* as a prominent resource of future anti-MRSA drugs. Front Microbiol 2018;9:2221.
 110. Rateb ME, Houssen WE, Arnold M, Abdelrahman MH, Deng H, Harrison WT, *et al.* Chaxamycins A-D, bioactive ansamycins from a hyperarid Desert *Streptomyces* sp. J Nat Prod 2011;74:1491-9.
 111. Gao W, Wu Z, Sun J, Ni X, Xia H. Modulation of kanamycin B and kanamycin A biosynthesis in *Streptomyces kanamyceticus* via metabolic engineering. PLoS One 2017;12:e0181971.
 112. Yu Z, Han C, Yu B, Zhao J, Yan Y, Huang S, *et al.* Taxonomic characterization, and secondary metabolite analysis of *Streptomyces triticiradicis* sp. nov.: A novel actinomycete with antifungal activity. Microorganisms 2020;8:77.
 113. Dulmage HT. The production of neomycin by *Streptomyces fradiae* in synthetic media. Appl Microbiol 1953;1:103-6.
 114. Fernández-Martínez LT, Borsetto C, Gomez-Escribano JP, Bibb MJ, Al-Bassam MM, Chandra G, *et al.* New insights into chloramphenicol biosynthesis in *Streptomyces venezuelae* ATCC 10712. Antimicrob Agents Chemother 2014;58:7441-50.
 115. Reynolds DM, Waksman SA. Grisein, an antibiotic produced by certain strains of *Streptomyces griseus*. J Bacteriol 1948;55:739-52.
 116. Anderson B, Hodgkin DC, Viswamitra MA. The structure of thiostrepton. Nature 1970;225:233-5.
 117. Maiti PK, Das S, Sahoo P, Mandal S. *Streptomyces* sp SM01 isolated from Indian soil produces a novel antibiotic picolinamycin effective against multi drug resistant bacterial strains. Sci Rep 2020;10:10092.
 118. Farver DK, Hedge DD, Lee SC. Ramoplanin: A lipoglycopeptide antibiotic. Ann Pharmacother 2005;39:863-8.
 119. Rahman H, Austin B, Mitchell W, Morris P, Jamieson D, Adams D, *et al.* Novel anti-infective compounds from marine bacteria. Mar Drugs 2010;8:498-518.
 120. Waksman SA, Geiger WB, Bugie E. Micromonosporin, an antibiotic substance from a little-known group of microorganisms. J Bacteriol 1947;53:355-7.
 121. Hiroshi S, Hideo O, Toshiaki H, Ikutoshi M, Kunio A, Mikio S. Thiolactomycin, a new antibiotic. J Antibiot 1982;35:396-400.
 122. Biffi G, Boretti G, Di Marco A, Pennella P. Metabolic behavior and chlortetracycline production by *Streptomyces aureofaciens* in liquid culture. Appl Microbiol 1954;2:288-93.
 123. Zygmunt WA. Oxytetracycline formation by *Streptomyces rimosus* in chemically defined media. Appl Microbiol 1961;9:502-7.
 124. Cai P, Kong F, Fink P, Ruppen ME, Williamson RT, Keiko T. Polyene antibiotics from *Streptomyces medicidicus*. J Nat Prod 2007;70:215-9.
 125. Grappel SF, Giovenella AJ, Phillips L, Pitkin DH, Nisbet LJ. Antimicrobial activity of aridicins, novel glycopeptide antibiotics with high and prolonged levels in blood. Antimicrob Agents Chemother 1985;28:660-2.
 126. Lucas X, Senger C, Erxleben A, Gruning BA, Doring K, Mosch J, *et al.* StreptomeDB: A resource for natural compounds isolated from *Streptomyces* species. Nucleic Acids Res 2013;41:D1130-6.
 127. Singh T, Wahla V. GC-MS analysis of antifungal compounds derived from soil actinobacteria. Int Res J Pharm 2018;9:81-4.
 128. Mojicevic M, D'Agostino PM, Nikodinovic-Runic J, Vasiljevic B, Gulder TA, Vojnovic S. Antifungal potential of bacterial rhizosphere isolates associated with three ethno-medicinal plants (poppy, chamomile, and nettle). Int Microbiol 2019;22:343-53.
 129. Andargie M, Li J. Antifungal activity against plant pathogens by compounds from *Streptoverticillium morookaense*. J Plant Pathol 2019;101:547-58.
 130. Jenifer JS, Michaelbabu M, Thirumalaikumar CL, Nisha SR, Uma G, Citarasu T. Antimicrobial potential of haloalkaliphilic *Nocardopsis* sp. AJ1 isolated from solar salterns in India. J Basic Microbiol 2019;59:288-301.
 131. Pattnaik S. Induction of bioactive compounds in a co-cultured strain of *Micromonospora echinospora* (DST-4) isolated from secluded Hirakud dyke soil. Indian J Biotechnol 2018;17:586-94.
 132. Mahdiyah D, Farida H, Riwanto I, Mustofa M, Wahjono H, Nugroho TL, *et al.* Screening of Indonesian peat soil bacteria producing antimicrobial compounds. Saudi J Biol Sci 2020;27:2604-11.
 133. Zhou L, Song C, Li Z, Kuipers OP. Antimicrobial activity screening of rhizosphere soil bacteria from tomato and genome-based analysis of their antimicrobial biosynthetic potential. BMC Genomics 2021;22:29.
 134. Malkov SV, Markelov VV, Polozov GY, Sobchuk LI, Zakharova NG, Barabanschikov BI, *et al.* Antitumor features of *Bacillus oligonitrophilus* KU-1 strain. J Microbiol Immunol Infect 2005;38:96-104.
 135. Abraham SA, Waterhouse DN, Mayer LD, Cullis PR, Madden TD, Bally MB. The Liposomal Formulation of Doxorubicin; 2005. p. 71-97. Available from: <https://www.linkinghub.elsevier.com/retrieve/pii/S0076687905910045> [Last accessed on 2022].
 136. Vijayabharathi R, Bruheim P, Andreassen T, Raja DS, Devi PB, Sathyabama S, *et al.* Assessment of resistomycin, as an anticancer compound isolated and characterized from *Streptomyces aurantiacus* AAA5. J Microbiol 2011;49:920-6.
 137. Long B, Rose W, Vyas D, Matson J, Forenza S. Discovery of antitumor indolocarbazoles: Rebeccamycin, NSC 655649, and fluorindolocarbazoles. Curr Med Chem Agents 2002;2:255-66.
 138. Hou J, Liu P, Qu H, Fu P, Wang Y, Wang Z, *et al.* Gilvocarcin HE:

- A new polyketide glycoside from *Streptomyces* sp. J Antibiot (Tokyo) 2012;65:523-6.
139. Balachandran C, Duraipandiyar V, Ignacimuthu S. Cytotoxic (A549) and antimicrobial effects of *Methylobacterium* sp. isolate (ERI-135) from Nilgiris forest soil, India. Asian Pac J Trop Biomed 2012;2:712-6.
 140. Egger C, Cannet C, Gérard C, Jarman E, Jarai G, Feige A, et al. Administration of bleomycin via the oropharyngeal aspiration route leads to sustained lung fibrosis in mice and rats as quantified by UTE-MRI and histology. PLoS One 2013;8:e63432.
 141. Segerman ZJ, Roy B, Hecht SM. Characterization of bleomycin-mediated cleavage of a hairpin DNA library. Biochemistry 2013;52:5315-27.
 142. Parthiban K, Vignesh V, Thirumurugan R. Characterization and *in vitro* studies on anticancer activity of exopolymer of *Bacillus thuringiensis* S13. Afr J Biotechnol 2014;13:2137-44.
 143. Kumar ML, Thippeswamy B, Raj PV. Cytotoxicity and anticancer studies of *Bacillus cereus* and *Bacillus pumilus* metabolites targeting human cancer cells. Appl Biochem Microbiol 2014;50:619-23.
 144. Xu DB, Ma M, Deng ZX, Hong K. Genotype-driven isolation of enterocin with novel bioactivities from mangrove-derived *Streptomyces qinglanensis* 172205. Appl Microbiol Biotechnol 2015;99:5825-32.
 145. Kaur S, Kaur S. Bacteriocins as potential anticancer agents. Front Pharmacol 2015;6:272.
 146. Izumikawa M, Takagi M, Shin-Ya K. JBIR-78 and JBIR-95: Phenylacetylated peptides isolated from *Kibdelosporangium* sp. AK-AA56. J Nat Prod 2012;75:280-4.
 147. Aliyu AB, Musa AM, Abdullahi MS, Ibrahim H, Oyewale AO. Phytochemical screening and antibacterial activities of *Vernonia ambigua*, *Vernonia blumeoides* and *Vernonia ocephala* (Asteraceae). Acta Pol Pharm 2011;68:67-73.
 148. He S, Feng Y, Ni J, Sun Y, Xue L, Feng Y, et al. Different responses of soil microbial metabolic activity to silver and iron oxide nanoparticles. Chemosphere 2016;147:195-202.
 149. Musa AM, Abbas G, Aliyu AB, Abdullahi MS, Akpulu IN. Phytochemical and antimicrobial screening of *Indigofera conferta* Gillett (*Papilionaceae*). Res J Med Plant 2008;2:74-8.
 150. Shen B, Du L, Sanchez C, Edwards DJ, Chen M, Murrell JM. The biosynthetic gene cluster for the anticancer drug bleomycin from *Streptomyces verticillus* ATCC15003 as a model for hybrid peptide-polyketide natural product biosynthesis. J Ind Microbiol Biotechnol 2001;27:378-85.
 151. Azman AS, Othman I, Fang CM, Chan KG, Goh BH, Lee LH. Antibacterial, anticancer and neuroprotective activities of rare actinobacteria from mangrove forest soils. Indian J Microbiol 2017;57:177-87.
 152. Malla S, Niraula NP, Singh B, Liou K, Sohng JK. Limitations in doxorubicin production from *Streptomyces peucetius*. Microbiol Res 2010;165:427-35.
 153. Hopp DC, Milanowski DJ, Rhea J, Jacobsen D, Rabenstein J, Smith C, et al. Citreamicins with potent gram-positive activity. J Nat Prod 2008;71:2032-5.
 154. Pei G, Dai H, Ren B, Liu X, Zhang L. Exploiting bioactive Enediynes from marine microbe based on activity and gene screening. Wei Sheng Wu Xue Bao 2010;50:472-7.
 155. Sugiura T, Ariyoshi Y, Negoro S, Nakamura S, Ikegami H, Takada M, et al. Phase I/II study of amrubicin, a novel 9-aminoanthracycline, in patients with advanced non-small-cell lung cancer. Invest New Drugs 2005;23:331-7.
 156. Gupta A, Chow M. Pimecrolimus: A review. J Eur Acad Dermatol Venereol 2003;17:493-503.
 157. Aftab U, Zechel DL, Sajid I. Antitumor compounds from *Streptomyces* sp. KML-2, isolated from Khewra salt mines, Pakistan. Biol Res 2015;48:58.
 158. Nguyen HT, Pokhrel AR, Nguyen CT, Pham VT, Dhakal D, Lim HN, et al. *Streptomyces* sp. VNI, a producer of diverse metabolites including non-natural furan-type anticancer compound. Sci Rep 2020;10:1756.
 159. Kim JW, Kwon Y, Bang S, Kwon HE, Park S, Lee Y, et al. Unusual bridged angucyclinones and potent anticancer compounds from *Streptomyces bulli* GJA1. Org Biomol Chem 2020;18:8443-9.
 160. Stutzman-Engwall KJ, Otten SL, Hutchinson CR. Regulation of secondary metabolism in *Streptomyces* spp. and overproduction of daunorubicin in *Streptomyces peucetius*. J Bacteriol 1992;174:144-54.
 161. Vijayakuma S, Saravanan V. *In vitro* Cytotoxicity and antimicrobial activity of biosurfactant produced by *Pseudomonas aeruginosa* Strain PB3A. Asian J Sci Res 2015;8:510-8.
 162. Kennedy RK, Naik PR, Veena V, Lakshmi BS, Lakshmi P, Krishna R, et al. 5-Methyl phenazine-1-carboxylic acid: A novel bioactive metabolite by a rhizosphere soil bacterium that exhibits potent antimicrobial and anticancer activities. Chem Biol Interact 2015;231:71-82.
 163. Ramasubburayan R, Sumathi S, Bercy DM, Immanuel G, Palavesam A. Antimicrobial, antioxidant and anticancer activities of mangrove associated bacterium *B. subtilis* subsp. *subtilis* RG. Biocatal Agric Biotechnol 2015;4:158-65.
 164. Kim YS, Balaraju K, Jeon YH. Biological characteristics of *Bacillus amyloliquefaciens* AK-0 and suppression of ginseng root rot caused by *Cylindrocarpon destructans*. J Appl Microbiol 2017;122:166-79.
 165. Ram Kumar A, Rajagopal K, Meenambiga SS, Kumaresan S. *In vitro* and *in silico* anti-breast cancer analysis of bioactive metabolites of *B. subtilis* isolated from soil. Saudi J Pathol Microbiol 2020;5:220-9.

How to cite this article:

Ram Kumar A, Kumaresan S. Antimicrobial and anticancer potential of soil bacterial metabolites - a comprehensive and updated review. J App Biol Biotech. 2023;11(5):1-11. DOI: 10.7324/JABB.2023.11501