

# Field treatment of three wheat varieties with *Trichoderma harzianum* bioagent to control *Anguina tritici*

Nawres Abdulelah Sadeq Alkuwaiti<sup>ID\*</sup>, Ammar Amjad Aish, Saad Tareq Abdulmalak, Tariq A. Kareem, Mohammed Mahmood Sulaiman

Plant Protection Department, College of Agricultural Engineering Sciences, University of Baghdad, Baghdad, Iraq.

## ARTICLE INFO

### Article history:

Received on: May 27, 2022

Accepted on: September 16, 2022

Available online: November 22, 2022

### Key words:

Phylogeny,  
Eco-friendly pesticide,  
Biocontrol,  
rDNA.

## ABSTRACT

A field experiment was performed to assess the susceptibility of *Triticum aestivum* L. var. Cham 6, Aras, and Baraka wheat local varieties, treated with *Trichoderma harzianum* Rifai bioagent, against seed gall infection. *Anguina tritici* identification was confirmed based on PCR amplification and sequence comparison of 18S small subunit ribosomal DNA. Sequence analysis showed *A. tritici* isolated from Wasit and Kirkuk provinces shared 100% maximum nucleotide sequence identity with the equivalent GenBank sequences from Mexico (AF363107) and India (JF826516), respectively, suggesting their common origin. Field experiment revealed that *T. aestivum* L. var. Aras was less susceptible to seed gall infection when scored 28.9% infectivity percent followed by Baraka and Cham 6 varieties which scored 29.6 and 33.1% infectivity, respectively. *T. harzianum* root watering treatment could decrease the disease incidence when scored 15.4, 17.5, and 19.7% compared to spraying treatment which scored 19.3, 20.8, and 22.4% infectivity, for Aras, Baraka, and Cham 6 varieties, respectively. Besides, root watering treatment reduced seed gall disease up to 46.71% compared to 33.22% for foliar spraying treatment. Thus, *T. harzianum* bioagent can be used as an ecofriendly nematicide alternative to control seed gall disease in Iraq.

## 1. INTRODUCTION

Wheat *Triticum aestivum* L. is a major grain crop grown worldwide within the family Poaceae [1]. It is thought this plant has originated from southeastern parts of Turkey 10,000 years BC [2]. Whereas in Iraq, bread wheat has been domesticated 9500 years BC [3]. Wheat is grown in Iraq in winter season. Based on 2020 statistics, the estimated Iraqi production and growing area of wheat were 6,238,392 tons and 2,143,421 ha, respectively [4]. About 46 different wheat varieties have been reported to be grown in Iraq [5]. Wheat is attacked by several pests including nematodes [1]. At least, four nematode groups infect wheat including cereal cyst, root knot, root lesion, and seed gall nematodes [5]. *Anguina tritici* (Steinbuch, 1799) Chitwood, 1935, or ear cockle nematode is a major plant pathogenic nematode infecting wheat causing serious losses worldwide [6]. It has been described for the 1<sup>st</sup> time by Needham since 1743 [1]. Seed gall disease has been reported in Iraq for the 1<sup>st</sup> time since 1921 [7] but the causal agent *A. tritici* was recently confirmed by molecular approach [8]. This disease can cause yield losses ranged 10.76–75% in wheat [9,10]. Limited studies conducted in Iraq were investigated

*A. tritici* control which were based on cultural, chemical, and biological methods [11–13]. Due to the limited molecular studies regarding seed gall disease in Iraq, this study, therefore, was initiated to identify the nematode causing seed gall disease in Wasit and Kirkuk provinces and its phylogenetic relatedness and to investigate the efficacy of the bioagent *Trichoderma harzianum* against seed gall disease on three wheat in the study area.

## 2. MATERIALS AND METHODS

### 2.1. Pathogenic Nematode Isolation and Inoculum Preparation

In 2021 growing season, gall samples were obtained from wheat fields located at Kirkuk and Wasit provinces in Iraq. Ten galls were soaked in 500 ml sterilized distilled water for 24 h at ambient temperature to release the second stage juveniles (J2). The number of J2 nematode was estimated using microscopic examination at 40X [14].

### 2.2. Molecular Conformation of the Pathogenic Nematode

Two seed galls from each location were crushed with liquid nitrogen and DNA extraction was performed using commercial DNA extraction kit (Bioneer, South Korea), following the manufacturer's instructions. PCR amplification was performed following [15], using AccuPower PCR PreMix commercial kit (Bioneer, South Korea) and ITS rRNA universal primers (rDNA2: TTGATTACGTCCCTGCCCTTT and rDNA1: ACGAGCCGAGTGATCCACCG) [16,17]. DNA

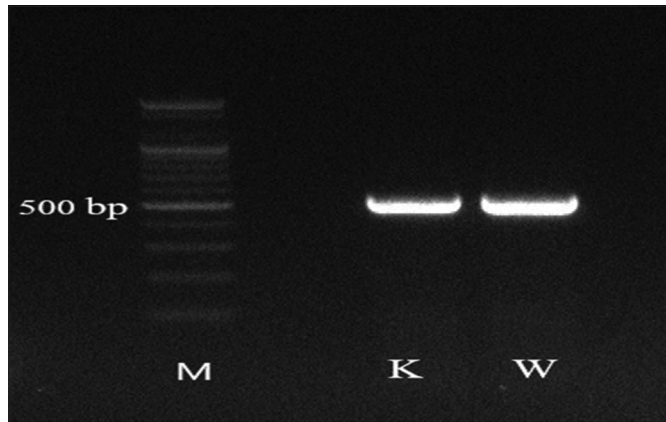
\*Corresponding Author:

Nawres Abdulelah Sadeq Alkuwaiti,  
Plant Protection Department, College of Agricultural Engineering Sciences,  
University of Baghdad, Baghdad, Iraq.  
E-mail: [n.alkuwaiti@coagri.uobaghdad.edu.iq](mailto:n.alkuwaiti@coagri.uobaghdad.edu.iq)

fragment amplified was visualized by ethidium bromide agarose gel electrophoresis [18]. PCR products were directly sequenced (Macrogen, South Korea). Sequences were analyzed using MEGA11 [19] and Sequence Demarcation Tool Version 1.2 (SDT v 1.2) [20] software packages.

### 2.3. Field Experiment

Local wheat varieties *T. aestivum* L. vars. Cham 6, Baraka, and Aras were tested against seed gall nematode infection. The experimental field was plotted into 1 m<sup>2</sup> areas with three replicates of each treatment.



**Figure 1:** Ethidium bromide stained gel pattern shows ~ 500 bp amplified by rDNA2/rDNA1 primer set from seed galls collected. K: Kirkuk sample, W: Wasit sample, and M: 100 bp DNA marker (Bioneer, S. Korea).

*T. harzianum* suspension (in the form of trichozone biopesticide, Al-Joud Company for Industry and Modern Agriculture, Iraq), was applied by foliar spraying or root watering at concentration 2 g/L. Nematode inoculum, adjusted to 10,000 J2 individuals' concentration, was applied at 1000 mL/m<sup>2</sup> rate, 25 days of seed sowing. All agricultural practices necessary for plant growth were followed, during the experiment. Infectivity percent and disease reduction (DR) were calculated using the following equation:

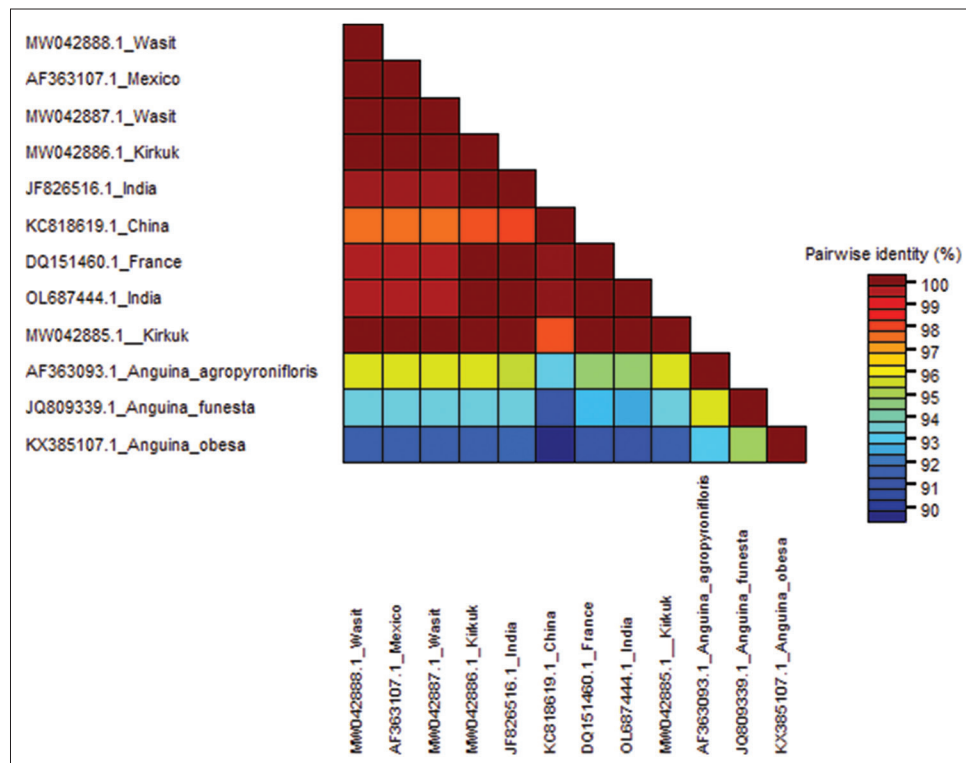
$$\text{Infectivity\%} = \left( \frac{\text{Number of infected seeds}}{\text{Total number of seeds}} \right) \times 100$$

$$\text{DR \%} = \left( \frac{\text{Number of seed galls in infected control} - \text{Number of seed galls in treatment}}{\text{Number of seeds galls in infected control}} \right) \times 100$$

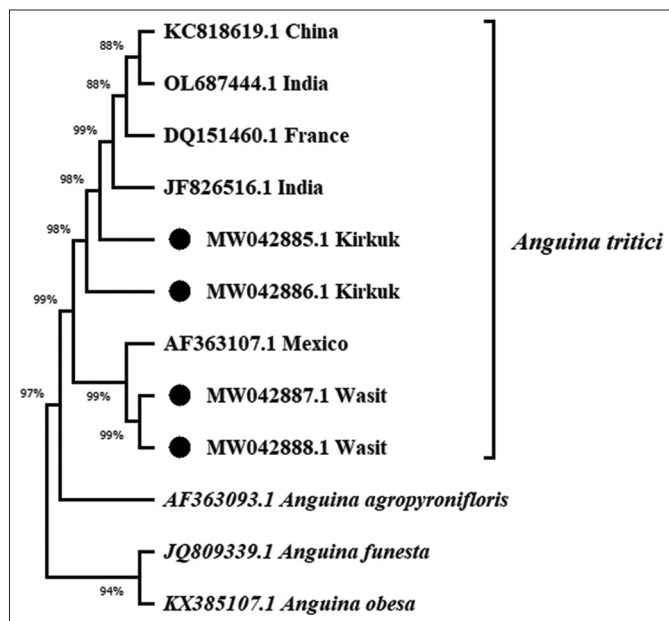
Microsoft Excel 2010 was used to analyze data calculated.

### 3. RESULTS AND DISCUSSION

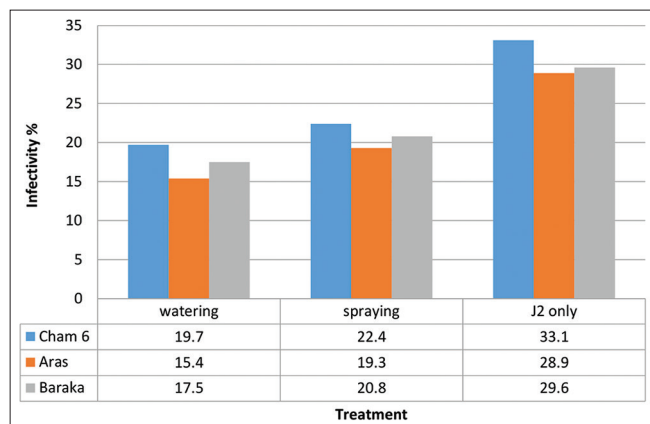
Sequence comparison confirmed DNA fragments amplified [Figure 1] were identical to 18S small subunit (SSU) ribosomal DNA genomic region of *A. tritici* retrieved from the NCBI [Figure 2]. *A. tritici* isolated from Wasit and Kirkuk scored 100% maximum nucleotide (nt) sequence identities to equivalent GenBank sequences from Mexico (AF363107) and India (JF826516), respectively, suggesting their common origin [Figure 2]. Neighbor-joining (NJ) phylogenetic tree, based on SSU nt sequences, confirmed the relatedness when grouped Wasit and Kirkuk sequences to relevant isolates from Mexico and India [Figure 2]. Despite the high identity, Wasit and Kirkuk isolates showed



**Figure 2:** Three color mode matrixes showing *A. tritici* identities, constructed from partial 18S SSU ribosomal DNA sequences of Kirkuk and Wasit and equivalent sequences from the GenBank. *A. agropyronifloris* Norton, 1965, *A. funesta* Price, Fisher and Kerr, 1979, and *A. obesa* Mobasseri, Pedram, Pourjam and Bertozzi, 2017, were used as out-group comparisons. Data were analyzed using Muscle algorithm. This matrix was generated using Sequence Demarcation Tool Version 1.2 (SDTv1.2) software package [20].



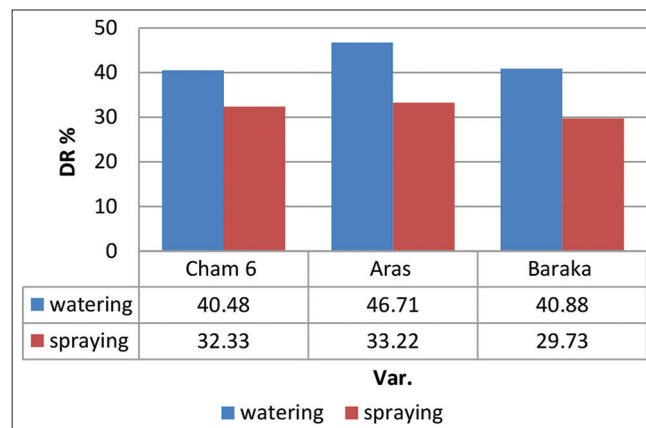
**Figure 3:** Neighbor-joining phylogenetic tree constructed from partial 18S SSU rDNA nucleotide sequences of *Anguina tritici* from Iraq (marked with ●) and equivalent sequences from the GenBank. *A. agropyronifloris* Norton, 1965, *A. funesta* Price, Fisher and Kerr, 1979, and *A. obesa* Mobasseri, Pedram, Pourjam and Bertozzi, 2017, were used as out-group comparisons. Data were analyzed using Maximum Composite Likelihood method. This tree was constructed by MEGA 11 software [19].



**Figure 4:** *T. harzianum* field treatments of three wheat varieties (Cham 6, Aras, and Baraka) against *A. tritici* using foliar spraying and root watering applications. Root watering treatments show the lowest infectivity percentages. J2: Represents untreated infarcted control. This bar chart was generated using Microsoft Excel 2010.

that they could be two variants when N-J phylogenetic tree separated them into two clades within *A. tritici* branch [Figure 3]. Thus, SSU could be useful molecular loci for identification, differentiation, and phylogeny reconstruction of *A. tritici* in Iraq [8,21].

This variation may be related to the selective pressure of many constraints favoring certain pathotypes over others in wheat growing area including varieties grown, alternative hosts available, and limited movements of nematode-infected plant materials, nationwide [22,23].



**Figure 5:** Disease reduction (DR) percentages resulted from *T. harzianum* field treatments of three wheat varieties (Cham 6, Aras, and Baraka) against *A. tritici* using foliar spraying and root watering applications. Root watering treatments show the lowest infectivity percentages. J2 only: Untreated infarcted control. This bar chart was generated using Microsoft Excel 2010.

Field experiment revealed that *T. harzianum* treatments could decrease infectivity percent and reduce seed gall disease in all three varieties compared to J2 only treatment. Root watering treatment showed high efficiency to control seed gall disease scoring 15.4% lowest infectivity [Figure 4] and 46.71% highest DR [Figure 5] in Aras variety, compared to foliar spraying. Root watering treatment with *T. harzianum* might minimize *A. tritici* infection much effectively than foliar spraying through direct interaction with J2 individuals, systemic induced resistance, and/or promoting or inducing plant growth [24,25]. This study confirmed the incidence of *A. tritici* in two Iraqi provinces, Kirkuk and Wasit. Based on phylogenetic relatedness and high identity percentage, these two geographical variants may have been introduced into Iraq through imported contaminated wheat seeds in the near past [26]. Except Saber-Beg, all wheat varieties are seed gall sensitive [27-29]. Aras, Baraka, and Cham6 wheat varieties showed to be sensitive to seed gall disease infection. Field experiment revealed that Aras wheat variety was less susceptible to seed call infection when scored 28.9% infectivity percent followed by Baraka and Cham 6 which scored 29.6 and 33.1% infectivity, respectively [Figure 4]. These three varieties are cultivated in Iraq due to their high yield quality and quantity compared to Saber-Beg. Bioagent treatment of desirable varieties can be the most efficient among others controlling methods as it offers a sustainable, eco-friendly alternative nematicide, to manage seed gall disease in Iraq [30]. Besides, it can enhance wheat production through promoting plant growth [24,25].

#### 4. CONCLUSIONS

This study confirmed the detection of *A. tritici* collected from two Iraqi provinces, Kirkuk and Wasit. Phylogenetic relatedness showed high identity percentage of the Iraqi seed gall nematode to the equivalent GenBank isolates from Mexico and India. Root treatment with *T. harzianum* was much effective than foliar treatment, against *A. tritici* infection, when decrease the infectivity percent of seed galls in all treatments. *T. aestivum* L. var. Aras was less susceptible to seed gall infection compared to Baraka and Cham 6 varieties. Bioagent treatment of desirable varieties can be the most efficient among others controlling methods as it offers a sustainable, eco-friendly alternative nematicide, to manage seed gall disease in Iraq.

## 5. ACKNOWLEDGMENT

Authors would like to thank the staff of the Seed Inspection and Certification directorate/Ministry of Agriculture and Miss Noor Raad Khuder and Mr. Ali Majid Jawad (Almusaib Bridge for Scientific and Lab Equipment) for their technical assistance.

## 6. AUTHORS' CONTRIBUTIONS

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the international committee of medical journal editors requirements/guidelines.

## 7. FUNDING

There is no funding to report.

## 8. CONFLICTS OF INTEREST

The authors report no financial or any other conflicts of interest in this work.

## 9. ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

## 10. DATA AVAILABILITY

The authors confirmed that all the relevant data were included in the article.

## 11. PUBLISHER'S NOTE

This journal remains neutral with regard to jurisdictional claims in published institutional affiliation.

## REFERENCES

- Skantar AM. *Anguina tritici* (Wheat Seed Gall Nematode). Invasive Species Compendium. Wallingford, UK: CABI; 2018. Available from: <https://www.cabi.org/isc/datasheet/5388> [Last accessed on 2022 Apr 03].
- Shewry PR. Wheat. J Exp Bot 2019;60:1537-53.
- Hirst KK. Wheat Domestication. Thought Co.; 2021. Available from: <https://www.thoughtco.com/wheat-domestication-the-history-170669> [Last accessed on 2022 May 16].
- FAOSTAT, Crops and Livestock Products. Available from: <https://www.fao.org/faostat/en/#data/QCL> [Last accessed on 2022 May 16].
- Wheat Atlas. Available from: <http://wheatatlas.org> [Last accessed on 2022 May 16].
- Dababat AA, Muminjanov H, Smiley R. Nematodes of small grain cereals current status and research. In: 5<sup>th</sup> International Cereal Nematode Initiative Workshop, 12–15 September 2015. Ankara, Turkey: Food and Agriculture Organization of the United Nations, Ankara, Turkey, 2015.
- Rao RS. A Preliminary List of Insect Pest of Iraq. Bombay: Times Press; 1921.
- Ami SN, Taher IE, Hussen FS, Ahmed AI. First molecular identification of wheat seed gall nematode *Anguina tritici* races parasitized on wheat in Iraq. Acta Univ Sapientiae Agric Environ 2019;11:5-15.
- Stephan ZA, Antoon BG. Biotypes of ear cockle nematode *Anguina tritici* in Iraq. Curr Nematol 1990;1:85-8.
- Ami SN, Taher IE. Survey, races identification and host range of wheat seed gall nematode *Anguina tritici* Duhok Province, Kurdistan region Iraq. J Agric Vet Sci 2014;7:44-8.
- Ami SN, Mustafa SA. Susceptibility of certain wheat and barley cultivars to seed gall nematode *Anguina tritici* (Steinbech, 1979) Filipjev, 1936. Mesopotamia J Agric 2012;40:217-23.
- Hassan AA, Hindi YK. Integrated control of *Anguina tritici* by some nematocides and *Trichoderma harzianum* isolated from wheat fields in Salah Aldin governorate. Tikrit J Agric Sci 2015;15:85-98.
- Al-Taie AH, Al-Zubaidi NK. Interaction efficiency of *Trichoderma* spp. and some plant extracts against ear-cockle disease. J Appl Biol Biotechnol 2022;10:102-7.
- Fattah FA, Al-Assas K. Histopathological comparison of galls induced by *Anguina tritici* with galls subsequently colonised by *Rathayibacter tritici* in wheat. Nematol Mediterr 2010;109:195-8.
- Szalanski AL, Sui DD, Harris TS, Powers TO. Identification of cyst nematodes of agronomic and regulatory concern by PCR-RFLP of ITS1. J Nematol 1997;29:255-67.
- Cherry T, Szalanski AL, Todd TC, Powers TO. The internal transcribed spacer region of *Belonolaimus* (Nemata: Belonolaimidae). J Nematol 1997;29:23-9.
- Vrain TC, Wakarchuk AC, Levesque AC, Hamilton RI. Intraspecific rDNA restriction fragment length polymorphism in the *Xiphinema americanum* group. Fundam Appl Nematol 1992;15:563-73.
- Sambrook JF, Russell D. Condensed Protocols: From Molecular Cloning: A Laboratory Manual. New York: Cold Spring Harbor Laboratory Press; 2006.
- Tamura K, Stecher G, Kumar S. MEGA11: Molecular evolutionary genetics analysis version 11. Mol Biol Evol 2021;38:3022-7.
- Muhire BM, Varsani A, Martin DP. SDT: A virus classification tool based on pairwise sequence alignment and identity calculation. PLoS One 2014;9:e108277.
- Carta LK, Li S. Improved 18S small subunit rDNA primers for problematic nematode amplification. J Nematol 2018;50:533-42.
- Lambert K, Bekal S. Introduction to plant-parasitic nematodes. Plant Health Instr 2002;10:1094-1218.
- Montarry J, Mimee B, Danchin EG, Koutsovoulos GD, Ste-Croix DT, Grenier E. Recent advances in population genomics of plant-parasitic nematodes. Phytopathology 2021;111:40-8.
- Poveda J, Abril-Urias P, Escobar C. Biological control of plant-parasitic nematodes by filamentous fungi inducers of resistance: *Trichoderma*, mycorrhizal and endophytic Fungi. Front Microbiol 2020;11:992.
- TariqJaveed M, Farooq T, Al-Hazmi AS, Hussain MD, Rehman AU. Role of *Trichoderma* as a biocontrol agent (BCA) of phytoparasitic nematodes and plant growth inducer. J Invertebr Pathol 2021;183:107626.
- Mackesy DZ, Sullivan M. CPHST Pest Datasheet for *Anguina tritici*. USDA-APHISPPQ-CPHST; 2016.
- AL-Jobouri M, AL-Jobouri A, AL-Qaissi E. Susceptibility of bread wheat cultivars to wheat gall nematode and a study some of the gene action of infection percentage. Kirkuk Univ J Sci Stud 2010;5:112-21.
- Qassem NE, AL-Taie HH, Thanoon AH. Screening of some varieties of wheat for infestation by the seed gall nematode *Anguina tritici*. Plant Cell Biotechnol Mol Biol 2021;22:94-105.
- AL-Jobouri J, AL-Jobouri R, Alsaedy H. Varietal and biological resistance for nematode no des of wheat *Anguina tritici* in genotypes

- of bread wheat *Triticum aestivum* L. Kirkuk Univ J Sci Stud 2021;11:96-111.
30. Błaszczyk L, Siwulski M, Sobieralski K, Lisiecka J, Jędryczka M. *Trichoderma* spp. application and prospects for use in organic farming and industry. J Plant Prot Res 2014;54:309-17.

**How to cite this article:**

Alkuwaiti NAS, Aish AA, Abdulmalak ST, Kareem TA, Sulaiman MM. Field treatment of three wheat varieties with *Trichoderma harzianum* bioagent to control *Anguina tritici*. J App Biol Biotech. 2023;11(1):200-204. DOI: 10.7324/JABB.2023.110128