

Multitargeted molecular docking study of phytochemicals on hepatocellular carcinoma

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ABSTRACT

Hepatocellular carcinoma (HCC) is the fourth major cause of death worldwide, with a global diminishing survival rate of 19%. Irrespective of the advanced therapeutic strategies against this carcinoma, it persists as one of the most challenging diseases. Moreover, the low efficacy of existing treatment stratagem using synthetic drugs against HCC has led to the urgent investigation of natural alternatives that can result in a more efficient treatment with fewer health side effects than their synthetic counterparts. In this study, a total of 1259 phytochemicals were docked against 25 potential HCC protein targets with the help of PyRx, a virtual screening tool software. The pharmacokinetics and drug-like properties of these chemicals were examined through SWISS ADME webserver. Based on their binding affinity against each protein target, only 250 ligands were shortlisted further for toxicity analysis using the web tools ADMETlab 2.0 and Protox II. In accordance with the bioavailability radar and pharmacokinetic profile analysis, only two non-toxic phytochemicals: Sorgolactone and Alectrol, emerged as the most befitting drug candidates against HCC protein targets 6HH1 and 1ZXM, respectively. The findings of this study suggest that these two phytochemicals can be explored and exploited further for their use as potential HCC drug candidates.

1. INTRODUCTION

Cancer is the leading cause of death and a significant impediment to living longer across the globe [1]. The World Health Organization 2019 estimate reveals that cancer is the first or second major cause of death in 112 countries [1,2]. The global burden of cancer incidence and death is on the rise; simply due to population increase and age, the worldwide burden of cancer has been anticipated to rise to 27.5 million new cancer cases and 16.3 million cancer deaths by 2040. In socioeconomically transitioning countries, the future burden will undoubtedly be more significant due to an increase in the incidence of cancer risk factors associated with unhealthy lifestyles such as smoking, consumption of alcohol and junk food, less physical activity, and low natality rates [1-5].

The primary liver cancer is the sixth most commonly diagnosed disease and the fourth major cause of cancer mortality globally.

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Furthermore, the prognosis of the disease is poor, with an estimated 5-year net survival rate of 19% [6,7]. Hepatocellular carcinoma (HCC) and intrahepatic cholangiocarcinoma (ICC) are the two most common histologic forms of the primary liver cancer. HCC comprises about 75% of the liver cancer cases, whereas ICC accounts for only about 12-15% of all the liver cancer cases throughout the world [6-9]. Hemangiocarcinoma accounts for 1% of all liver cancers, whereas secondary liver cancer or liver metastasis accounts for the rest of the liver cancer cases [10]. HCC develops in hepatocytes, usually due to oxidative stress, inflammation, and underlying liver disease, whereas ICC develops in the cholangiocytes that border the intrahepatic bile duct [8,11]. Despite breakthroughs in cancer treatment, liver cancer remains one of the most challenging malignancies to treat. Surgery, local destructive therapy, and liver transplantation have all been shown to be curative for people with the early HCC, but recurrence of HCC remains a hindrance in the treatment of the disease with an incidence rate of 70% or more. HCC is frequently discovered at an advanced stage, and many patients in these stages are ineligible for curative treatments. Furthermore, standard chemotherapy has low efficacy and provides less benefit in terms of survival [12]. Thus, several species of microbes, plants, animals, and marine animals are now proving to be promising sources for discovering new therapeutic candidates [13].

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With the discovery and development of different natural substances such as cytotoxic podophyllotoxins, vincristine, vinca alkaloids, and vinblastine in the 1950s, the quest for anticancer agents from plant sources began [14]. The successful clinical use of cancer chemotherapeutic drugs derived from plants for nearly half a century has piqued interest for further research [14-16]. With advancements in technology, scientists have improved natural product drug discovery in an industrial setting. Several natural anticancer drugs have been launched in the market, and several drugs originating from various organisms are currently being tested in cancer-related clinical trials [14-17]. There are numerous routes involved in the development of cancer. Blocking one or more components of a specific pathway involved in cancer formation is a common strategy for developing anticancer drugs [9].

The molecular docking approach may be used to represent the atomic level interaction between a molecule and a protein, allowing us to define the molecule's behavior in target protein binding sites as well as to elucidate key biochemical processes [18-20]. The molecules identified through docking are more likely to advance to the next stage of the drug development process. Nowadays, *in vitro* and *in silico* techniques are commonly utilized to examine the pharmacokinetic features of the chemical entities, and molecular modeling is widely used for optimizing leads in drug development.

The present study investigates chemicals and phytochemicals as potential drug inhibitors against liver cancer through *in silico* approach. To examine the potential of phytochemicals, different protein targets were selected. These targets included apoptotic proteins, transcription factors, serine/threonine protein kinases, growth factor receptors, mitogen-activated protein kinases (MAPK), and cell division protein kinase. We have performed a molecular docking study of ligands against all these proteins. Further, information from this study could be valuable in exploring a new compound, which could be very useful in developing a drug against HCC.

2. MATERIALS AND METHODS

2.1. Target Protein Retrieval

A total of 25 potent protein targets are involved in hepatocarcinoma, ranging from MAPK, nuclear receptor, apoptotic proteins, transcription factors, topoisomerases, epidermal growth factor receptor, cyclin dependent kinases (CDK), serine/threonine protein kinases, and various other proteins were selected for docking against the selected ligands [Table 1].

PDB IDs for these proteins were obtained from protein data bank (PDB) [21], and their respective PDB format files were downloaded. To obtain processed protein structures free from any duplicate chains, these PDB files were opened using a 3D molecular visualization software, the BIOVIA Discovery Studio Visualizer (https://discover.3ds. com/discovery-studio-visualizer-download) and then the prebound ligand groups, heteroatoms and water molecules, and any undesired interference from the groups mentioned above were eliminated by their deletion.

2.2. Drug Preparation

For this study, a total of 1259 potential phytochemical drugs which might exhibit anticancer activity against HCC were retrieved from the Dr. Duke's Phytochemical and Ethnobotanical Databases (http://phytochem.nal.usda.gov/). These databases aid in an extensive search for herbal and therapeutic plants and their chemical, bioac-

tive, ethnobotanical characteristics using common, and scientific botanical names. The PubChem database (https://pubchem.ncbi. nlm.nih.gov/) was used to obtain the 2D structure SDF files for each drug.

2.3. Energy Minimization of Docking Entities

The energy of processed proteins was then minimized using the Chimera software version 1.15 [22] through its Surface/Binding Analysis Tool for Dock prep, which facilitated the addition of required hydrogen atoms and charges acquired from Dunbrack Library at relevant positions in protein structure. The ligands finalized for docking analysis were initially energy minimized through the optimization algorithm by first loading their 2D structure SDF files on the PyRx – Virtual Screening Tool Software [23] and then converted to PDBQT format through its Open Babel tool. At this point, both the docking entities were ready for the next step.

2.4. Docking Methodology

In the molecular docking domain, to obtain a desired stabilized drug-receptor complex, it is essential to erase any unnecessary initial interactions of the two molecules with other prebound molecules to facilitate binding of the ligand at a competitive site on the receptor macromolecule. Before setting up the docking process, the energy minimized file of each target protein was first converted to a macromolecule. The docking operation was then launched using the Vina Wizard Control from the PyRx – Virtual Screening Tool Software by setting the Vina Search Space at its maximum as well as setting up the Vina Run exhaustiveness at value eight and finally continuing with the procedure that displays results in the form of binding affinity (kcal/mol) and root mean square deviation (RMSD) lower and upper bound values for each run up until the selected run value.

2.5. Visualization and Analysis of Docking Results

Out of all the RMSD values, only the highest scores were screened. Therefore, the ten topmost ligands displaying the highest binding affinity scores against each protein were selected for further analysis. The complex files of the drugs with their ligands were visualized in the form of a 2D drug-receptor complex in the BIOVIA Discovery Studio Visualizer. The structure of the desired ligand as per the binding affinity score and target-protein complex for each of the 25 proteins was visualized for Van der Waals, conventional H-bond, π -anion, π -cation, amide π -stacked, alkyl and π -alkyl, π -sigma, π - π T shaped, Carbon H-bond, π -Sulfur, π - π Stacked, Unfavorable Donor-Donor, and Unfavorable Acceptor-Acceptor interactions.

2.6. Bioavailability Radar and Pharmacokinetic (SWISS ADME) Profile Analysis

The canonical SMILES of each of the selected ligands with a high binding affinity against their targets were obtained from the simple search engine of the Indian Medicinal Plants, Phytochemistry and Therapeutics database [24]. Further using this data, the Pharmacokinetic analysis report was obtained from the SWISS ADME web tool (http://www.swissadme.ch/), a division of the web tools group provided by the Swiss Drug Design platform supported by the Swiss Institute Bioinformatics that act an as efficient computer-aided drug designing platform. The above-mentioned web tool platform evaluated the drug-likeness as per the bioavailability Table 1: List of protein targets for docking analysis.

PDB ID	Protein description	Protein function	Protein	Resolution (A°)	References
1CM8	Proline-directed serine/threonine kinases	MAP kinases are dominant factors in the processes that govern embryogenesis, cell differentiation, proliferation, and cell death	MAP kinase	2.4	[21]
1ERR	Nuclear transcription factors	Responsible for reproductive, skeletal, cardiovascular, and central nervous systems			[22]
1GFW	Cysteine proteases	Major executioner caspase during the demolition phase of apoptosis	Caspase 3	2.80	[9]
1JUN	Transcription factor	The factor plays a role in cellular processes, which includes cell proliferation, survival, and transformation of cell	AP-1	Highest	[23]
1NFK	Transcription factor	Plays a role in cell proliferation and differentiation, inflammatory and immune response, cell survival and apoptosis, cellular stress reactions, and tumorigenesis	NF-κB p50 subunit	2.3	[22]
1P22	Nuclear effector of canonical Wnt signaling in the nucleus	Tissue homeostasis, cell renewal, and regeneration	β-catenin	2.95	[23]
1ZXM	Complex multifunctional enzymes	Modulation of topological state, chromosome segregation, and chromatin condensation	Human topoisomerase IIa	1.87	[24]
2AR9	Cysteine proteases	The caspase-9 function is vital for apoptosis during normal development of the central nervous system	Caspase 9	2.8	[9]
2CLS	Rho GTPase	Human RND1 expression is dysregulated in tumors and is involved in oncogenesis GTPase in the active GTP bound state		2.31	[19]
2E2B	Tyrosine-protein kinase	Cell growth and survival such as cytoskeleton c-Abl kinase remodeling in response to extracellular stimuli, cell domain in complex motility and adhesion, receptor endocytosis, autophagy, with INNO-406 DNA damage response, and apoptosis		2.20	[20]
2HY8	Serine/Threonine protein kinases	Plays a role in cell cycle, cell motility, apoptosis, and cell survival	PAK1	2	[22]
2HZI	Tyrosine kinase	Responsible for cell differentiation, cell division, cell adhesion, and stress response			[25]
20H4	Belongs to PDGF supergene family	Signals for endothelial cell growth and survival (angiogenic)	Vascular EGFR-2	2.05	[9]
20JI	Protein kinase	Proliferation, differentiation, transcription, and cellular motility	ERK2	2.60	[22]
3EZV	Serine/Threonine protein kinase	Role in the G1/S transition, the initiation of DNA synthesis, and the regulation of the exit from the S phase	CDK-2	1.99	[24]
3UPI	RNA-dependent RNA polymerase	HCV RNA replication	NS5B	2.00	[26]
4AQ3	BLC-2 family members regulate apoptosis	Family (such as BCL-2 and BCL-XL) inhibit apoptosis; meanwhile, others (such as BAX and BAK) promote cell death	BCL-2	2.40	[22]
4JSX	Serine/threonine protein kinase that belongs to Phosphatidylinositol-3-kinase related kinase superfamily	mTORC1 controls protein synthesis, cell growth, MTOR proliferation, autophagy, cell metabolism, and stress responses, whereas mTORC2 seems to regulate cell survival and polarity		3.50	[22]
4LQM	Family of receptor tyrosine kinases	Role in cell proliferation, survival, differentiation, and migration	EGFR	2.50	[9]
4Y7R	Oncoprotein transcription factor	Cell growth, proliferation, loss of differentiation, and apoptosis	c-Myc	1.898	[23]
4Y83	Proto-oncogenic kinase, which belongs to the MAP3K family	COT contributed to G1 phase progression of T-lymphocytes, perhaps via regulation of IL-2 secretion	COT kinase	2.89	[27]

Table 1: (Continued)

PDB ID	Protein description	Protein function	Protein	Resolution (A°)	References
5G4N	Tumour-suppressor protein	Role of p53 in the regulation of glycolysis and autophagy, the repair of genotoxic damage, cell survival and regulation of oxidative stress, invasion and motility, cellular senescence, angiogenesis, differentiation, and bone remodeling	p53	1.35	[23]
5LA9	Heterodimeric transcription factor	Role in body's response to low oxygen concentrations, or hypoxia	HIF-1α	2.81	[23]
5CT7	Kinase	Role in cellular proliferation, differentiation, and survival	BRAF in complex with RAF265	3.17	[20]
6HH1	Receptor tyrosine kinase	Intracellular signaling and the mutated form of c-Kit plays a crucial role in the occurrence of some cancers	c-Kit with allosteric inhibitor 3G8	2.25	[20]

 $NF-\kappa B$: Nuclear factor Kappa-B, $ER-\alpha$: Estrogen receptor- α , CDK-2: Cyclin-dependent kinase-2, PAK1: p21-activated kinase, NS5B: Non-structural protein 5B, BCL-2: B-cell lymphoma 2, MTOR: Mammalian target of rapamycin, EGFR: Endothelial growth factor receptor, COT: Cancer Osaka thyroid, HIF-1 α : Hypoxia-inducible factor 1 α , PDGF: Platelet-derived growth factor, and ERK2: Extracellular-signal-regulated kinase 2, HCV: Hepatitis B virus

radar image and data obtained for each ligand as per Lipinski's rule of five.

2.7. Computational Toxicity Prediction

Ligands following Lipinski's rule of five were further evaluated to predict numerous toxicity parameters using the webtools ADMETlab 2.0 [25] and Protox II [26]. ADMETlab 2.0 aids in screening the drug molecules for Absorption, Distribution, Metabolism, Excretion, and Toxicological characteristics (ADMET), whereas Protox II, a virtual in silico laboratory, facilitated in mimicking its conventional counterpart and thereby delivered toxicity prediction of small molecules. Each ligand's canonical smile was used to study its toxicity profile. Protox II helps determine the median lethal dose 50 (LD50) value in mg/kg along with toxicity class prediction. The toxicity classes are subdivided into six subtypes from Class I to VI, wherein Class I $(LD50 \le 5)$ depicts an LD50 value that can be lethal if the drug is swallowed, and Class VI (LD50 > 5000) represents a non-toxic LD50 value. Furthermore, the tool also provides a toxicity model report which consists of variables such as Hepatotoxicity, Carcinogenicity, Immunotoxicity, Mutagenicity, and Cytotoxicity.

ADMET laboratory 2.0 precisely evaluates the toxic potential of the molecule through toxicity variables such as human ether-a-go-go generelated blockers, human-hepatotoxicity, drug-Induced liver injury, ames test for mutagenicity, eye irritation, rat oral acute toxicity, FDAMDD (maximum recommended daily dose), skin sensitization, carcinogenicity, eye corrosion, respiratory toxicity, and environmental toxicity.

3. RESULTS AND DISCUSSION

Liver cancer is among the most prevalent malignant tumors in the world. The Global Cancer Statistics 2018 indicates that it ranks seventh and third in terms of global incidence and mortality, respectively, among all known cancer types. In all, 840,000 new cases and 780,000 deaths associated with HCC were reported in 2018 alone [27,28]. Chronic alcoholism, hepatitis B virus and hepatitis C virus infections, non-alcoholic fatty liver disease, and several other factors such as drinking water pollution, microcystins, and aflatoxin B1 also play a significant role in the incidence of HCC across the globe [29-32]. HCC is difficult to diagnose and cure; thus, using a phytochemical for developing a cure for this disease would be interesting and intriguing. In this study, we have investigated the binding of different phytochemicals to 25 protein targets associated with liver cancer using computational docking approaches.

A total of 1259 phytochemicals were chosen based on their ability to inhibit the proteins associated with HCC. These phytochemicals were retrieved from Dr Duke's phytochemical and ethnobotanical databases. The phytochemicals chosen were organic compounds such as fatty acids, carbohydrates, benzene and its derivatives, chromones, polyphenols, and amino acids.

3.1. Molecular Docking

Molecular docking can investigate the interaction of a protein with ligands. It aims to find the potent drug by binding the correct ligand poses into the protein binding pockets [Table 2]. It elucidates whether the protein-ligand complex improves or inhibits the protein activity. It filters the best protein-ligand pair according to their binding scores so that the most potent drug candidate can be used for further testing. Molecular docking was carried out with the selected 25 proteins and the 1259 bioavailable ligands. A total of 250 ligands consisting of the top ten ligands of each protein ranked on the basis of their binding energy values are presented in Table 3. The higher binding affinity scores indicate that the ligands show a perfect fit into the protein cavity; the higher binding affinity can be attributed to the presence of hydrogen bonds in the complex. The π -cation bonds balance the negative atoms such as chlorine and fluorine in the drug, whereas the π -sigma bond introduces the charges responsible for the intercalation of the drug in the binding sites of the protein.

Hydrophobic ligand interaction in the receptor-binding pocket is enhanced by bonds such as π -alkyl and alkyl [32]. The highest number of hydrogen bonds was observed in the case of ligands that displayed higher binding affinity against the protein.

3.2. Evaluation of Pharmacokinetic and Pharmacological Properties

The Lipinski's rule of five was used to determine the drug-likeness of the phytochemicals examined in the study Table B of the supplementary data. The rule states that the drug-like compound must have LogP values ranging between 0 and 5, hydrogen bond donors <5, hydrogen bonds acceptors <10, and the drug's molecular weight should not be more than 500 Da [33]. Thus, the rule is based on the molecular weight, the presence of hydrogen donor-acceptor bonds, and the drug's lipophilicity. These factors are used to evaluate the drug's absorption, penetration, metabolism, and excretion. The interaction of the membrane with the drug is affected when the compound violates the rule of five; poor absorption and permeation of the drug are more

 Table 2: Grid box parameters selected for target enzyme, based on the binding site residues.

Target proteins	Active site residues	Grid box parameters	
1CM8	GLU-181, MET-182, MET-201, TYR-203, GLY-184, TRP-200, ARG-73, ARG-176, LYS-69, ARG-70, ARG152, ASP-153, VAL-186, VAL-187, ARG-192, ARG-180, HIS-231, ASP-230, PRO-110, MET-109, ILE-87, ALA-54, PHE-111, MET-112, LYS-56, LEU-170, ASP-171, VAL-41, ALA-40, ALA-160, ASN-158, GLY-157, LYS-155, LYS-118, VAL-33, THR-114, ASP-115	26.51*54.90*25.77	83.52*86.38*95.40
1ERR	MET-343, PHE-404, LEU-346, LEU-349, THR-347, ARG-394, LEU-391, ALA-350, ASP-351, GLU-353, LEU-354, LEU-539, LEU-536, VAL-376, HIS-377, LEU-378, GLY-457, SER-456, LEU-379, GLU-380, ALA-382, TRP-383, LEU-387, LEU-384, GLU-385, MET-388, LEU-391, ARG-394, LEU-525, GLY-521, MET-522, SER-518, ASN-519, ARG-515, HIS-524, LEU-428, ILE-424, PHE-425, MET-421, GLY-420	50.09*37.17*68.42	69.16*60.29*59.30
1GFW	GLU-125, GLY-122, HIS-121, SER-120, MET-61, CYS-163, ALA-162, TYR-204, SER-205, TRP-206, PHE-256, ARG-207, SER-65, SER-209	26.71*22.60*37.12	57.21*54.90*53.24
1JUN	GLY-274, CYS-273, CYS-273, GLY-274, GLY-275, ARG-276, ILE-277, ALA-278, LEU-280, GLU-281, ARG-276, ILE-277, LEU-280, GLU-281, ALA-278, GLY-275	2.40*1.04*1.63	23.87*67.83*22.32
1NFK	GLY-52, PRO-68, GLY-66, GLY-65, SER-78, SER-72, GLU-73, GLY-77	15.42*20.85*11.10	59*82*59
1P22	ARG-431, ALA-39, GLY-38, GLY-432, GLY-408, ALA-391, ALA-392, LEU-351,	17.92*32.00*-2.12	88*85*52
1ZXM	LYS-168, PHE-142, ALA-167, GLY-166, TYR-165, ILE-141, GLN-376, THR-147, GLY-164, ASN-163, SER-148, TYR-34, SER-149, ARG-162, ASN-150, GLY-161, ILE-125, LYS-378, THR-215, ALA-92, ARG-98, ASN-120, ASN-91, ASP-94, ASN-95, LYS-123, GLY-160, GLU-87, GLY-124	63.65*7.81*18.65	54.05*49.76*75.15
2AR9	PRO-357, ASP-356, ARG-355, TRP-354, SER-353	15.73*41.00*0.96	101*129*147
2CLS	LEU-129, PHE-170, LYS-126, ALA-169, SER-168, MET-29, LYS-26, GLY-25, CYS-24, TYR-38, ASP-21, THR-68, GLY-70, SER-69, SER-71, THR-45, VAL-22, PRO-39, GLU-40, THR-41, PRO-44, TYR-42, VAL-43,	14.88*-17.11*25.90	71*64*73
2E2B	ARG-362, ILE-360, PHE-359, HIS-361, LEU-354, VAL-289, ILE-293, GLU-286, MET-290, VAL-379, ASP-381, ALA-380, LEU-298, PHE-382, VAL-299, LEU-370, TYR-253, GLY-321, MET-318, THR-319, LEU-248, THR-315, ILE-313, GLU-316, LYS-271, PHE-317, VAL-256, ALA-269, VAL-270	13.64*71.86*39.60	59.28*90.41*91.77
2НҮ8	GLY-349, GLY-350, ALA-348, LEU-347, SER-351, LEU-396, VAL-328, ASP-393, ASN-394, THR-406, ASP-407, GLU-315, ALA-297, TYR-346, GLU-345, MET-344, ILE-276, GLY-277, VAL-284, GLN-278, ARG-299, GLY-279, SER-281	11*79*14	53*58*44
2HZI	GLU-286, PHE-382, MET-290, ASP-381, ALA-380, ILE-313, VAL-299, THR-315, GLU-316, TYR-253, LYS-271, VAL-256, LEU-370, VAL-270, THR-315, ALA-269, GLU-316, GLY-249, ASN-322, LEU-248, GLY-321, TYR-320, THR-319	19.34*12.33*14.72	40*50*60
20H4	ASP-1056, ASP-1054, PRO-1055, LYS-869, ALA-842, LYS-1053, PRO-837, ARG-840, GLY-841, VAL-1058, ILE-1051, VAL-846, GLY-839, ARG-1049, ASP-1050, ARG-1059, LEU-838, LYS-866, LEU-1047, ALA-1048, ALA-864, GLY-1046, ASP-1062, GLU-883, PHE-1045, VAL-914, ILE-886, ASP-1044, PHE-916, LEU-887, GLU-915, GLY-920, LYS-918, CYS-1043, ASN-1031, ARG-1030, CYS-917, PHE-919, VAL-897, LEU-1033, HIS-1024, ILE-890, ILE-1042, VAL-896, LEU-1017	5.13*37.98*23.09	64*44*68
20JI	ALA-33, GLY-32, GLU-31, GLY-30, ILE-29, GLY-35, VAL-37, LYS-52, LYS-112, GLU-69, ASP-165, ALA-50, ASN-152, ASP-109, THR-108, GLN-103, CYS-164, ASP-104, LEU-105, LEU-154, MET-106	-4.33*8.39*46.93	64*44*68
3EZV	PHE-146, ASP-145, ALA-144, VAL-64, PHE-80, GLU-81, LYS-33, VAL-18, ALA-31, LEU-134, PHE-82, LEU-83, GLN-131, ILE-10, ASP-86, GLN-85, HIS-84	2.23*30.54*21.73	64*44*68
3UPI	ASP-318, ASP-319, GLY-317, ASN-316, SER-367, CYS-366, SER-368, ARG-386, SER-556, TYR-555, GLY-449, TYR-448, PHE-193, GLN-446, ILE-447, GLY-410, ASN-411, MET-414, PRO-197, TYR-415, ARG-200, LEU-384	59.11*-8.23*-24.60	48.42*65.43*56.95
4AQ3	MET-74, ASP-70, PHE-71, TYR-67, ARG-66, ASP-62, PHE-63, ALA-59, TYR-161, LEU-160, PHE-112, VAL-92, LEU-96, GLU-95, THR-91, ALA-108, VAL-107, ARG-105, TRP-103, ASN-102, PHE-157	-13.93*14.15*-10.15	40.6*34.74*39.13
4JSX	TRP-2239, VAL-2240, CYS-2243, LEU-2185, TYR-2225, ILE-2237, ASP-2244, ILE-2163, PRO-2169, LYS-2187, SER-2165, THR-2245, MET-2345, ILE-2356, SER-2342, ASP-2195, ASP-2357, GLU-2190	69.64*-12.59*-49.70	89.77*104.60*117.47
4LQM	GLN-791, THR-790, GLU-762, LEU-792, MET-793, ILE-789, LEU-788, ALA-743, ILE-744, LYS-745, VAL-726, LEU-718, THR-854, LEU-844, ASP-855, GLY-796, CYS-797, ARG-841, ASP-800	23.20*24.98*21.05	65.73*109.12*112.22

 Table 2: (Continued)

Target proteins	Active site residues	Grid box parameters	
4Y7R	CYS-134, PHE-133, SER-175, ASP-92, SER-91, CYS-261, TYR-260, ILE-266, PHE-263, SER-50, SER-306, SER-49, ILE-264, ILE-305	13.02*23.79*8.34	53.34*54.94*37.99
4Y83	LEU-134, LYS-133, TRP-132, GLU-217, ILE-144, GLY-213, SER-214, VAL-152, ARG-146, SER-257, GLY-210, ALA-165, LYS-167, ALA-209, VAL-260, ASN-258, GLU-208, MET-207, VAL-269, ASP-270, ALA-191	17.52*-34.78*-13.22	40.96*45.69*49.23
5G4N	VAL-157, GLU-221, CYS-220, LEU-257, THR-230, PRO-222, PRO-223, LEU-145, CYS-229, ASP-228, TRP-146, PRO-151, PHE-109, VAL-147, THR-150	130.5*91.75*-29.77	40.96*45.69*49.23
5LA9	TRP-389, MET-299, THR-387, THR-325, PRO-402, ASP-315, ALA-301, ILE-327, ALA-385, ASP-254, PHE-366, ARG-252, HIS-313, TYR-303, TYR-310, TYR-329, ARG-383, HIS-374, VAL-376, LEU-343	-24.02*23.32*16.59	54.03*47.37*39.90
5CT7	GLY-534, GLU-533, ILE-463, PHE-583, CYS-532, TRP-531, ALA-481, VAL-471, GLN-530, VAL-482, PHE-595, THR-529, LYS-483, ILE-592, LEU-514, GLY-593, ASP-594, ILE-527, ILE-513, LEU-567, HIS-574, THR-508, LEU-505, ILE-572, VAL-504, GLU-501	-15.73*10.38*0.88	64*44*68
6HH1	LEU-783, CYS-788, ILE-789, HIS-790, ILE-808, ILE-653, LEU-647, LEU-799, CYS-809, VAL-654, ASP-810, VAL-643, LEU-644, CYS-673, PHE-811, GLU-671, GLU-640, TYR-670, ALA-621, LEU-595, VAL-603, LYS-623	57.54*38.47*14.68	54.58*50.44*57.58

Table 3: Top ten ligands showcasing highest binding affinity scores against each protein candidate.

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
1CM8	Apiforol	443638	-11.9
	Adrenosterone	223997	-10.5
	Alstonine	441979	-9.7
	Shinjulactone A	460537	-9.7
	Hippeastrine	441594	9.5
	Kurchinicin	9873539	-9.4
	3'Methoxycoumestrol	44151023	-9.4
	Coumestrol	5281707	-9.3
	Carbamazepine	2554	-9.2
	Uzarigenin	92760	9.2
ERR	Venalstonine	426061	-10.1
	Sorgolactone	5281395	-9.7
	Butin	92775	9.5
	Epicatechin	255538	-9.5
	Naringenin	439246	-9.5
	7,3',4'Trihydroxyflavone	5322065	-9.4
	Enterolactone	10685477	-9.4
	Garbanzol	442410	9.3
	Gibberellin A51	443458	-9.3
	Apiforol	443638	-9.3
GFW	Pamoic acid	8546	-9.1
	Uzarigenin	92760	-8.9
	Digitoxigenin	4369270	-8.9
	4Hydroxystrychnine	211181	-8.8
	Jatrophatrione	5281372	-8.8
	Strychnine	441071	-8.7
	CHEMBL3612189	5245667	-8.7
	Pseudostrychnine	21723446	-8.6
	Diterpene II (lactone)	339816	-8.5
	Brucine	442021	-8.5

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
JUN	Psoralenol	5320772	-6.4
	Corylidin	5316096	-6.2
	Isoarboreol	21722929	-6.2
	Alectrol	5281353	-6.1
	Gmelanone	21722946	-6.1
	Pamoic acid	8546	-6
	Jatrophatrione	5281372	-5.9
	Indirubin	5359405	-5.9
	Protopine	4970	-5.8
	β-colubrine	251890	-5.8
INFK	Amarolide	460539	-8.5
	Corylidin	5316096	-8.4
	2Hydroxy3methoxystrychnine	21680050	-8.4
	Pseudostrychnine	21723446	-8.2
	Protopine	4970	-8.1
	4Hydroxystrychnine	211181	-8.1
	β-colubrine	251890	-8.1
	Pamoic acid	8546	-8
	Stylopine	440583	-8
	Sorgolactone	5281395	-8
1P22	Pseudostrychnine	21723446	-8.8
	Strychnine	441071	-8.6
	Isoarboreol	21722929	-8.6
	Uzarigenin	92760	-8.4
	Digitoxigenin	4369270	-8.4
	Gmelanone	21722946	-8.4
	4Hydroxystrychnine	211181	-8.3
	CHEMBL3612189	5245667	-8.3
	2Hydroxy3methoxystrychnine	21680050	-8.2
	10-Hydroxycamptothecin	97226	-8.2
1ZXM	Gmelanone	21722946	-11.4
	(+) Teframidine	436140	-11.2
	Stylopine	440583	-11.1
	Seneciphylline	5281750	-11.1
	Diterpene II (lactone)	339816	-10.9
	Alectrol	5281353	-10.8
	Ent7alpha, 12betadihydroxy16kauren19,6betaolide	5088389	-10.7
	Fumaritine	21627986	-10.6
	19hydroxytabersonine	443325	-10.2
	Gibberellin A51	443458	-10.2
2AR9	10-hydroxycamptothecin	97226	-9.4
	Berberine	2353	-9
	Corytuberine	160500	-8.6
	Cleomiscosin A	442510	-8.6
	Berberastine	442180	-8.4
	Allamcin	5477870	-8.3
	4'Hydroxychalcone 4'glucoside	23144947	-8.2

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
	Protopine	4970	-8.1
	Pseudoyohimbine	251562	-8
	Yohimbine	8969	-7.9
2CLS	Isoarboreol	21722929	-10.6
	Gmelanone	21722946	-10.4
	Cleomiscosin A	442510	-10.3
	Bavachromanol	21722946 442510 5321790 9976812 5320772 5322065 5281220 5281614 246330 5281707 363863 2353 5359405 442667	-10.2
	(+) Neoolivil	9976812	-9.8
	Psoralenol	5320772	-9.7
	7,3',4'Trihydroxyflavone	5322065	-9.7
	Aureusidin	5281220	-9.6
	Fisetin	5281614	-9.6
	Dihydrofisetin	246330	-9.5
2E2B	Coumestrol	5281707	-10.9
	CHEBI: 73029	363863	-10.8
	Berberine	2353	-10.8
	Indirubin	5359405	-10.7
	Anhydroglycinol	442667	-10.6
	Benzo[a] phenoxazin-9-ylidene (dimethyl) azanium	81507	-10.5
	Hematoxylin	442514	-10.5
	2hydroxychrysophanol	442759	-10.5
	Jutamansinone	759294	-10.5
	Equol	91469	-10.4
2HY8	Methoxystrychinine	21723445	-10.3
	β-colubrine	251890	-10.1
	Strychnine	441071	-10.1
	CHEMBL3612189	5245667	-10.1
	Pseudostrychnine	21723446	-10
	4Hydroxystrychnine	211181	-9.7
	Brucine	442021	-9.7
	Dehydroevodiamines	9817839	-9.7
	Protopine	4970	-9.6
	10-hydroxycamptothecin	97226	-9.6
2HZI	Indirubin	5359405	-11.8
	Benzo[a] phenoxazin-9-ylidene (dimethyl) azanium	81507	-11.4
	CHEMBL3612189	5245667	-11.4
	Medicagol	5319322	-11.4
	Paulownin	3084131	-11.3
	Psoralenol	5320772	-11.2
	Carbamazepine	2554	-11.1
	CHEBI: 73029	363863	-11.1
	6Methoxypulcherrimin	44260092	-11.1
	Pulviatide	90471471	-11
2OH4	Fisetin	5281614	-9.7
	Butin	92775	-9.6
	Orobol	5281801	-9.6

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
	Pseudobaptigenin	5281805	-9.6
	7,3',4'Trihydroxyflavone	5322065	-9.6
	Santal	9926336	-9.6
	Naringenin	439246	-9.5
	Garbanzol	442410	-9.5
	Kaempferol	5280863	-9.5
	Genistein	5280961	-9.5
20JI	Pseudostrychnine	21723446	-10
	Shinjulactone A	460537	-9.9
	CHEMBL3612189	5245667	-9.9
	Corylidin	5316096	-9.8
	Strychnine	441071	-9.7
	Ent7α,12βDihydroxy16kauren19,6βolide	5088389	-9.4
	Molephantinin	5281485	-9.4
	4Hydroxystrychnine	211181	-9.3
	β-colubrine	251890	-9.3
	Precalyone	324879	-9.3
3EZV	Dehydroevodiamines	9817839	-10.3
	Corylidin	5316096	-10.2
	Rosmaquinone B	46883407	-9.9
	CHEMBL3612189	5245667	-9.8
	Bavachromanol	5321790	-9.8
	Pamoic acid	8546	-9.6
	4Hydroxystrychnine	211181	-9.6
	Aureusidin	5281220	-9.6
	Sulfuretin	5281295	-9.6
	(E) 7Hydroxy3 (4hydroxybenzylidene) chroman4one	44443280	-9.6
3UPI	Corylidin	5316096	-10.2
	Protopine	4970	-9.8
	Shinjulactone A	460537	-9.6
	Pamoic acid	8546	-9.5
	Medicagol	5319322	-9.5
	β-colubrine	251890	-9.4
	4Hydroxystrychnine	211181	-9.2
	Riddelliine	5281744	-9.2
	Granisetron	5284566	-9.2
	AC1L80TG	430162	-9.1
4AQ3	Protopine	4970	-8.7
C.	(+) Teframidine	436140	-8.6
	Jatrophatrione	5281372	-8.4
	Gmelanone	21722946	-8.3
	Methoxystrychinine	21723445	-8.3
	CHEMBL3612189	5245667	-8.2
	Centratherin	44409502	-8.2
	Brucine	44409302	-8.1
	Isopinnatal	3035834	-8.1
	Isophiliatai	23144947	-8.1

 Table 3: (Continued)

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
4JSX	Isoarboreol	21722929	-10.9
	Digitoxigenin	4369270	-10.7
	Uzarigenin	92760	-10.6
	Psoralenol	5320772	-10.5
	Stylopine	440583	-10.4
	(-)-8-oxotetrahydropalmatine	49769861	-10.2
	Pseudoyohimbine	251562	-9.9
	Medicagol	5319322	-9.9
	Dihydrobaicalein	9816931	-9.9
	Gibberellin A35	21596345	-9.9
4LQM	Strychnine	441071	-9.7
	Pseudostrychnine	21723446	-9.7
	Protopine	4970	-9.5
	Sorgolactone	5281395	-9.4
	Diosbulbin G	21723242	-9.4
	β-colubrine	251890	-9.2
	Cathenamine	443361	-9.2
	CHEMBL3612189	5245667	-9.2
	10-Hydroxycamptothecin	97226	-9.2
	Uzarigenin	92760	-9.1
4Y7R	Pamoic acid	8546	-10
	Pseudostrychnine	21723446	-9.8
	Shinjulactone A	460537	-9.7
	Granisetron	5284566	-9.7
	2Hydroxy3methoxystrychnine	21680050	-9.4
	4Hydroxystrychnine	211181	-9.2
	(+) Adlumine	442155	-9.1
	Amarolide	460539	-9.1
	Methoxystrychinine	21723445	-9.1
	Gmelanone	21722946	-9.1
4Y83	Pamoic acid	8546	-10.8
1105	Texalin	473253	-10.7
	(E) 7Hydroxy3 (4hydroxybenzylidene) chroman4one	44443280	-10.7
	Persicogenin	320054	-10.5
	Aureusidin	5281220	-10.5
	7,3',4'Trihydroxyflavone	5322065	-10.5
	10-Hydroxycamptothecin	97226	-10.5
	Calycosin	5280448	-10.4
	Naringenin	439246	-10.3
	Apiforol	443638	-10.3
5G4N	Medicagol	5319322	-8.8
	CHEMBL3612189	5245667	-8.5
	Coumestrol	5281707	-8.3
	CHEBI: 73029	363863	-8.2
	Hippeastrine	441594	-8.1
	Dehydroevodiamines	9817839	-8.1

(Contd...)

Table 3: (Continued)

Protein	Ligand Chemical Name	PubChem ID	Binding affinity (-kcal/mol)
	Protopine	4970	-8
	Pamoic acid	8546	-8
	Stylopine	440583	-8
	6Hydroxyrubiadin	5319801	-8
5LA9	Pamoic acid	8546	-8.5
	Strychnine	441071	-8.4
	Shinjulactone A	460537	-8.4
	4Hydroxystrychnine	211181	-8.3
	β-colubrine	251890	-8.3
	Corylidin	5316096	-8.3
	Akuammiline	5367018	-8.3
	2Hydroxy3methoxystrychnine	21680050	-8.3
	Viroallosecurinine	908416	-8.2
	Methoxystrychinine	21723445	-8.2
5CT7	Pseudobaptigenin	5281805	-10.5
	Santal	9926336	-10.2
	1,9pyrazoloanthrone	8515	-10
	Genistein	5280961	-10
	Thalidomide	5426	-9.9
	2'hydroxydihydrodaidzein	440047	-9.9
	2'Hydroxydaidzein	5280520	-9.9
	Baptigenin	9965663	-9.9
	Chebi: 73029	363863	-9.8
	3',4',7trihydroxyisoflavone	5284648	-9.8
6HH1	CHEMBL3612189	5245667	-10.1
	Stylopine	440583	-10
	Sorgolactone	5281395	-9.9
	Enterolactone	10685477	-9.9
	Cathenamine	443361	-9.8
	Dehydroevodiamines	9817839	-9.8
	Paulownin	3084131	-9.7
	Corylidin	5316096	-9.6
	Psoralenol	5320772	-9.6
	Protopine	4970	-9.5

expected when the rules are violated [34]. Lipinski's rule of five was used to determine the drug-likeness of the 250 phytochemicals that have higher binding affinity against the selected proteins. Compounds that violated one or more of Lipinski's rules were excluded from the study, and the remaining compounds were further investigated to test their toxicity. Furthermore, hydrogen bonds play a key role in drug designing as they are involved in drug permeation, metabolism, and absorption. The bond type and the number of bonds present in each protein-ligand complex are displayed in Table C of the supplementary data.

3.3. Bioavailability Radar and Toxicity Prediction

Following the molecular docking, pharmacokinetic, and pharmacological analysis, the selected compounds were, further, investigated for their toxicity profile. Bioavailability radar was utilized to determine the drug-likeness of the phytochemical against the respective protein. The tool determines the bioavailability radar of the given phytochemical based on six properties: saturation, size, solubility, flexibility, lipophilicity, and polarity. The saturation, that is, a fraction of carbons in the sp³ hybridization, should not be <0.25, and the molecular weight should range from 150 to 500 g/Mol. Solubility, that is, LogS should not be higher than 6, the flexibility should not be more than nine rotatable bonds, the lipophilicity, that is, XLOGP3 should be between -0.7 and +5.0, and the polarity, that is, TPSA must be between 20 and 130 Å² [35].

Oral drug formulations are widely used and more convenient to administer than the other drugs; they also have fewer sterility restrictions and are cost-effective. However, most of the new compounds have low water solubility, making oral drug consumption less suitable as a model for drug administration, thus making it a significant problem for drug developers. The high lipophilicity of the compounds also impacts the drug's therapeutic effect. These problems should be tackled to make the drug bioavailable. The compounds investigated against HCC were found to be orally bioavailable. Computational methods and approaches can be used to test a drug's safety during the developmental process. These have proven to have more advantageous over *in vitro* and *in vivo* studies [36]. The toxicity analysis of the shortlisted molecules was carried out using the PROTOX -II webtool. The parameters used for evaluating the compounds were: Cytotoxicity, Carcinogenicity, Hepatotoxicity, Immunotoxicity, Mutagenicity, Predicted LD50, and Toxicity (Oral toxicity).

Considering the bioavailability radars (Table B of the Supplementary data) of the best 250 phytochemicals that displayed the highest binding affinity scores against their respective receptors, only 20 ligands were found to fit in the pink colored optimal range of the bioavailability radar [Figure 1]. In addition, they were also found to be inactive for hepatotoxicity while being either active or inactive for other toxicity parameters, as well as displayed an appreciable number of hydrogen bonds with the respective HCC protein targets [Table 4].

Out of all the ligands displayed in Table 4, Enterolactone, Adrenosterone, Kurchinicin, Gmelanone, Protopine, Stylopine, Corylidin, and Shinjulactone A belonged to toxicity class IV ($300 < LD50 \le 2000$), indicating their harmfulness if swallowed. On the other hand, only three ligands, namely, Jatrophatrione, Amarolide, and Psoralenol, fell in the toxicity Class V ($2000 < LD50 \le 5000$), indicating their possibility of being toxic if swallowed. Finally, two phytochemical drugs, Sorgolactone and Alectrol, were observed to have the same LD50 value of 5105 mg/kg body weight. Such LD50 value >5000 mg/kg body weight indicates a Class VI toxicity level that is nontoxic. Based on these toxicity parameters, the phytochemicals Sorgolactone and Alectrol were considered as ideal drug targets against their respective HCC protein targets.

3.4. Selection of Drug Candidate for 6HH1

Based on the molecular docking, bioavailability radar, and toxicity profile analysis of the ligands, Sorgolactone was an ideal drug candidate against the tyrosine kinase receptor. Sorgolactone belongs to the group of strigolactones (SLs), which consist of terpenoid-derived endogenous plant hormones [37]. SLs have been known for three main physiological processes in plants: first, they aid in the germination of parasitic microbes which reside in the roots of the host plants of the genus *Striga*; second, they help in plant identification by the mutual association forming fungi; and finally, these phytochemicals are well known to prevent branching especially at plants' the stem terminal region. Sorgolactone plays a key role as a plant growth regulator involved in developing several parts of the plant such as stems, leaves, flowers, and the ripening of fruits.

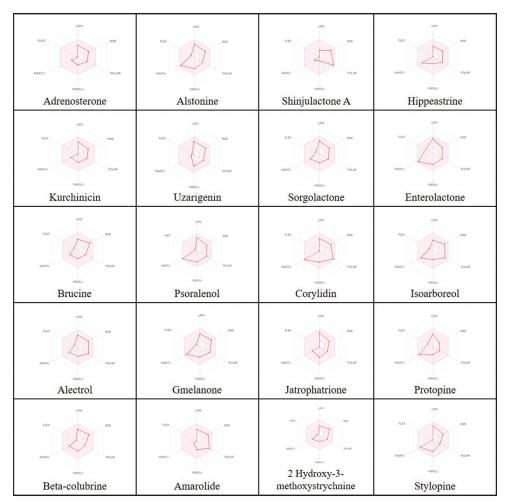


Figure 1: Bioavailability Radar plot of each shortlisted ligand.

Sorgolactone exhibited binding energy of -9.9 kcal/mol against the tyrosine kinase receptor 6HH1. Figure 2 depicts seven different types of bonds formed by sorgolactone with the kinase receptor: the conventional hydrogen bond, the alkyl, the π -alkyl, the π -anion, the π -cation, the π -sigma, and the π -sulfur bond. The 2D representation of the protein-ligand complex revealed that the ligand formed one conventional hydrogen bond on the 6HH1 chain A amino acid ASP-810; seven alkyl bonds were formed on LYS-623, VAL-603, CYS-788, LEU-644, and VAL-643; two π -alkyl bonds were formed with VAL-654 and LEU-644; one π -cation bond was formed with LYS-623, whereas two π -anion bonds were formed with ASP-810; one π -sulfur bond was created with the receptor amino acid CYS-809, and one π -sigma bond was formed with VAL-643.

3.5. Selection of a Drug Candidate for 1ZXM

Alectrol was selected as an appropriate candidate against the HCC protein target 1ZXM due to its high binding affinity, its accordance with Lipinski's rule of five, and its non-toxic nature as compared to other drug candidates. Alectrol belongs to the class of strigolactone, a class of cartenoid-derived molecules known for controlling plants' growth, development, adaptation, and abiotic stress regulation [38]. The binding affinity score of Alectrol against the protein molecule 1ZXM, a complex multifunctional enzyme Human topoisomerase IIa, was observed to be –10.8 kcal/mol and displays altogether ten bonds with the protein target. Figure 3 exhibits six conventional hydrogen bonds formed by electron on the receptor chain A amino acids, namely, GLY-164, ARG-162, SER-148, and SER-149, whereas two

Table 4: Toxicity profile of ligands.

Ligands	Toxicity Class	LD50 (mg/kg)	Hepato- toxicity	Carcino- genicity	Immuno- toxicity	Muta- genicity	Cyto-toxicity	Conventional hydrogen bonds
Sorgolactone	6	5105	Inactive	Active	Active	Inactive	Inactive	1
Alectrol	6	5105	Inactive	Inactive	Active	Inactive	Inactive	6
Jatrophatrione	5	4600	Inactive	Active	Active	Inactive	Inactive	2
Amarolide	5	3900	Inactive	Inactive	Active	Inactive	Inactive	2
Psoralenol	5	2875	Inactive	Inactive	Active	Inactive	Inactive	1
Enterolactone	4	2000	Inactive	Inactive	Active	Inactive	Inactive	2
Adrenosterone	4	1870	Inactive	Active	Active	Inactive	Inactive	1
Kurchinicin	4	1680	Inactive	Active	Active	Inactive	Active	2
Isoarboreol	3	1500	Inactive	Inactive	Active	Inactive	Inactive	3
Gmelanone	4	1500	Inactive	Active	Active	Inactive	Inactive	4
Protopine	4	940	Inactive	Active	Active	Active	Inactive	1
Stylopine	4	940	Inactive	Active	Active	Inactive	Active	2
Corylidin	4	832	Inactive	Active	Active	Active	Inactive	3
Shinjulactone A	4	590	Inactive	Inactive	Active	Inactive	Active	5
Hippeastrine	3	230	Inactive	Active	Active	Inactive	Inactive	1
Ac119c1h (synonyms: alstonine; senegasaponins b)	3	215	Inactive	Inactive	Active	Inactive	Inactive	1
Brucine	3	150	Inactive	Active	Inactive	Inactive	Inactive	2
Beta-colubrine	3	150	Inactive	Inactive	Active	Inactive	Inactive	1
2 Hydroxy-3-methoxystrychnine	3	150	Inactive	Inactive	Active	Inactive	Active	2
Uzarigenin	2	26	Inactive	Inactive	Active	Inactive	Inactive	3

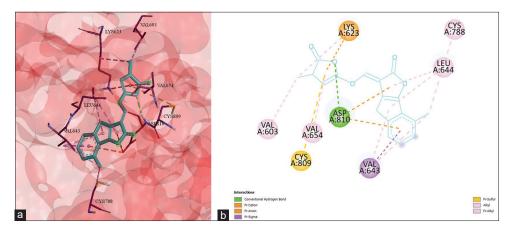


Figure 2: (a) 3D and (b) 2D representation of Sorgolactone interacting with the binding pocket of TYR kinase receptor 6HH1.

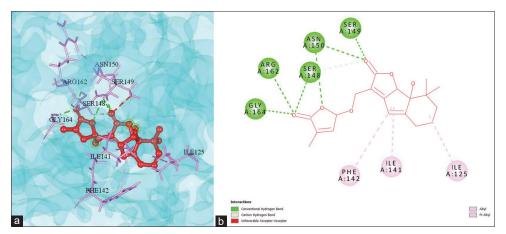


Figure 3: (a) 3D and (b) 2D representation of Alectrol interacting with the binding pocket of the complex multifunctional enzyme 1ZXM.

hydrogen bond linkages were formed with ASN-150. The drug was also observed to form a Carbon hydrogen bond with the amino acid LYS-168; a π -alkyl bond was formed with PHE-142, and one alkyl bond was created with the amino acids ILE-141 and ILE-125 of target protein 1ZXM.

4. CONCLUSION

Cancer has been a long thriving mortality threat to humanity for decades. Out of all the carcinoma types encountered to date, primary liver cancer continues to be the most frequently diagnosed disease and ranks fourth in terms of cancer mortality worldwide. Considering the threat posed by this carcinoma type, the search for effective drug candidates has been on the rise. As opposed to the synthetic drugs available for cancer treatment, naturally occurring chemicals known as phytochemicals are being investigated for their possible applications as anticancer agents. In our study, 1259 phytochemicals were docked against 25 HCC targets resulting in 250 probable drug candidates. With further screening using pharmacokinetic and pharmacological parameters and toxicity profiling of each possible drug candidate, only two non-toxic phytochemicals were identified as the most appropriate drug candidates against their respective carcinoma receptors.

Both the phytochemicals, Sorgolactone and Alectrol, belong to the class of SLs, which are widely known as plant hormones capable of restricting branching in plants [39]. Although our current study findings indicate that Sorgolactone and Alectrol have been identified as potential drug targets against the HCC protein targets 6HH1 and 1ZXM, respectively, both the phytochemicals were found to be active for immunotoxicity. In contrast, the former drug candidate was active for carcinogenicity [Table 4]. A probable solution to tackle this problem can be the introduction of a slight structural modification that can improve the chances of these drug candidates becoming less toxic. A review in the past has reported strigolactone analogues as potential anticancer agents that can affect the cancerous cells viability rates, promote apoptosis, and thereby lead to cell death at micromolar concentrations [40]. Another recent study conducted by Prandi et al. [41] reported the antiproliferative effects of SLs analogues on both solid and non-solid cancer cells lines such as prostate, colon, lung, melanoma, osteosarcoma, and leukemic cell line while being non-toxic to normal cell lines. In addition, the findings of the study report that SLs analogues have had an anti-proliferative effect on HCC-HepG2 cell line. Despite these studies, there has been no report till date towards the in silico and in vivo identification of Sorgolactone and Alectrol, both belonging to SLs, and their candidacy as potential

drug target. Therefore, the results of this study make an attempt to shed light on the potential of these two phytochemicals as probable drugs for therapeutic treatment against HCC.

5. AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work, and approved it for publication.

6. FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

7. CONFLICTS OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflicts of interest.

8. ETHICS APPROVALS

This study does not involve experiments on animals or human subjects.

9. DATA AVAILABILITY STATEMENT

All data generated or analyzed during this study are included in this published article (and its supplementary information files).

10. PUBLISHER'S NOTE

This journal remains neutral with regard to jurisdictional claims in published institutional affiliation.

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