



# Vitamin C pretreatment mitigates the genotoxic effects of sodium arsenite in human lymphocytes *in vitro*

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## ABSTRACT

The present study was aimed to evaluate Vitamin C (VC) as a potent natural antioxidant to reduce the genotoxic effects of sodium arsenite in human lymphocytes. The study was design to assess the role of VC (10 µg/ml), upon sodium arsenite induced *in vitro* genotoxicity by using alkaline comet assay technique. Lymphocytes were incubated for 3h at 37°C in RPMI-1640 medium with different concentrations of sodium arsenite (12.5 µg/ml, 25 µg/ml, 50 µg/ml and 100 µg/ml) and also in combined treatment of sodium arsenite with VC (100 µg/ml + 10 µg/ml). Our findings suggest that treatment with sodium arsenite increased the primary DNA damage and pretreatment with VC reduces the genotoxic effects of sodium arsenite effectively and efficiently at the highest concentration *in vitro*, proving its mitigating effect.

## 1. INTRODUCTION

Arsenic is well known water contaminant raising several health issues throughout the world [1]. Exposure to arsenic occurs via the ingestion, inhalation, dermal contact and the parenteral route to some extent. Humans can be exposed to arsenic through the intake of air, food and water [2]. Arsenic has been considered as a potent toxicant and carcinogenic to humans [3]. Long-term exposure to arsenic-contaminated water causes a wide range of adverse health effects, including skin pigmentations, keratosis, vascular diseases, and conjunctivitis in the eyes, neuropathy, lung diseases and different types of cancer [4]. Earlier works on arsenic clearly indicate the genotoxic potential of arsenic both *in vivo* and *in vitro* [5, 6]. *In vitro* exposure of arsenic caused primary DNA damage in human peripheral blood lymphocytes [7]. The toxicity of arsenic depends on its chemical state. Arsenic can occur in the environment in several oxidation states (-3, 0, +3 and +5) but in natural water mostly found in organic form as oxyanions of trivalent arsenite (AS III) or pentavalent arsenic (ASV) [8]. Inorganic arsenic in its trivalent form is more toxic than pentavalent arsenic. Sodium arsenite is pentavalent form of inorganic arsenic species. Most studies show that inorganic arsenic is genotoxic and cytotoxic *in vitro* and

*in vivo* [9]. The property of VC to increase the activity of glutathione (GSH) reductase, superoxide dismutase (SOD) and catalase (CAT) in swiss albino mice has been implicated to possibly play a protective role against chemical toxicity [10]. VC is the most widely used antioxidant and prevents oxidative damage to the cell membrane induced by radicals [11].

VC can protect the body from damage by ROS that can be generated during normal metabolism and other exposure like toxins and carcinogens. Previous work has reported the use of VC to combat arsenic-induced genotoxicity [12, 13]. The present study was designed to confirm the genotoxic potential of inorganic arsenic *in vitro* and vitamin c was selected as the most potential natural antioxidant to prevail over arsenite induced genotoxicity

## 2. MATERIALS AND METHODS

### 2.1 Chemicals

Sodium arsenite (CAS No. 7784-465), Phosphate buffered saline (PBS), trizma base were obtained from Hi-Media Ltd., Mumbai, India. Histopaque was purchased from Sigma-Aldrich Co. (USA). Vitamin C (CAS No.50-81-7), Normal melting point (NMP) agarose, low melting point (LMP) agarose, di-sodium salt of ethylene diamine tetra acetic acid (EDTA), Tris buffer, ethidium bromide (EtBr), thiobarbituric acid (TBA), trichloroacetic acid (TCA), Triton X-100, DMSO, sodium hydroxide, sodium chloride, acetic acid and methanol were obtained from Sisco Research Laboratories Pvt. Ltd. (Mumbai, India).

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## 2.2 Sample Collection and Isolation of lymphocytes

Heparinised vacutainers were used for collection of human peripheral blood from healthy volunteers. Lymphocytes were isolated from fresh blood by using Histopaque [14]. The cells were washed with PBS and resuspended in RPMI-1640 media at a concentration of  $10^6$  cell  $\text{mL}^{-1}$  for further use. This experimental study was approved by the Ethics Committee of Calcutta University.

## 2.3 Experimental design

The study included seven groups and presented in Table 1. In all the groups the exposure time for test chemical was 3 hr.

## 2.4 Treatment of the lymphocytes

Based on the results of the viability test 100  $\mu\text{g/ml}$  the concentration of sodium arsenite was selected as the  $\text{LD}_{50}$ ; as at this concentration approximately 50% of the cells died during the 3 hours of treatment. Therefore further all investigations were carried out using a concentration of 100  $\mu\text{g/ml}$  sodium arsenite.

Incubation of freshly isolated lymphocytes was done at 37 °C for 3h in RPMI-1640 media with different concentrations of sodium arsenite (12.5  $\mu\text{g/ml}$ , 25  $\mu\text{g/ml}$ , 50  $\mu\text{g/ml}$  and 100  $\mu\text{g/ml}$ ). Following treatment, the lymphocytes were processed for detection of DNA damage by alkaline comet assay.

## 2.5 Trypan blue dye exclusion method

Cytotoxicity was determined by trypan blue dye exclusion method [15]. Viable cells were counted in a Neubauer chamber under a light microscope.

## 2.6 Comet assay *in vitro*

The DNA damage were studied by alkaline comet assay [16, 17]. Slides were prepared in triplicates per concentration. Slides were immersed in cold lysis solution at  $\text{pH}$  10. The lysis solution consisted of 2.5M NaCl, 100mM  $\text{Na}_2\text{EDTA}$ , 10mM Trizma base, 1% TritonX 100, 10% DMSO and kept at 4°C for 60 min. After lysis the DNA was allowed to unwind in the electrophoresis buffer (300 mM NaOH: 1mM  $\text{Na}_2\text{EDTA}$  at  $\text{pH}$  13.5) for 20 min. This was followed by electrophoresis conducted at a constant voltage of 25V and 300 mA at 4°C. Slides were neutralized in 0.4M Tris ( $\text{pH}$  7.5) for 5 min and finally rinsed in water. Each experiment was repeated thrice.

## 2.7 Scoring of slides

EtBr (20  $\mu\text{g/mL}^{-1}$ ) was used for staining the slides and rinsing the slides in water to wash off excess stain. Slides were scored using image analysis system (Kinetic imaging; Andor Technology, Nottingham, UK) attached to a fluorescence microscope (Leica, Wetzlar, Germany). The microscope was connected to a computer through a charge coupled device (CCD) camera to transport images to software (Komet 5.5) for analysis. The final magnification was 100X. Among the comet parameters we report the percent tail DNA, tail extent moment and olive tail moment. This will give us a clear indication of the extent of DNA

damage induced by the test chemical. Images of 150 (50X3) cells per concentration were analysed for human lymphocytes. The median values of each concentration with respect to the comet parameter were calculated.

## 2.8 Statistical analysis

One-way analysis of variance (ANOVA) was used to analyze the % Tail DNA, Tail Extent Moment and Olive Tail Moment data at different concentration treatment. All the data were expressed as the mean  $\pm$  standard deviation (SD). Pair wise comparison between the study groups was done using Tukey's multiple comparison tests. The level of significance was taken as  $P < 0.05$ .

## 3. Result and Discussion

### 3.1 Cell Viability by Trypan blue dye exclusion method

Human lymphocytes were treated with different concentrations of Sodium arsenite (12.5, 25, 50 and 100 $\mu\text{g/ml}$ ), VC (10 $\mu\text{g/ml}$ ) and highest dose of sodium arsenite with VC (100 $\mu\text{g/ml}$  + 10 $\mu\text{g/ml}$ ) as described earlier (Table 1). The results of cell viability assays after incubation of lymphocytes with the sodium arsenite at different concentrations are displayed in Table 2. The cutoff point was 70% [18]. Viability was measured both before and after treatment and range of viable cells was within 75–90% in all experiments. The viability varied between 95% in control set to 81–86% at different treatment (12.5, 25, 50 and 100 $\mu\text{g/ml}$ ) of sodium arsenite. The cell viability decreased with increase in the concentration of sodium arsenite as compared with control ( $P < 0.001$ ). In the combined treatment of VC at concentration of 10  $\mu\text{g/ml}$  and highest concentration of sodium arsenite 100  $\mu\text{g/ml}$  in the culture increased the cell viability ( $P < 0.001$ ) and proving the ameliorative effect of VC presented in (Table 2).

**Table 1:** Description of the experimental design.

Experimental groups	Treatment	Doses
Group I	Control	
Group II	Sodium arsenite (NA1)	NA1 (12.5 $\mu\text{g/ml}$ )
Group III	Sodium arsenite (NA2)	NA2 (25 $\mu\text{g/ml}$ )
Group IV	Sodium arsenite (NA3)	NA3 (50 $\mu\text{g/ml}$ )
Group V	Sodium arsenite (NA4)	NA4 (100 $\mu\text{g/ml}$ )
Group VI	Vitamin C (VC)	VC (10 $\mu\text{g/ml}$ )
Group VII	Sodium arsenite + Vitamin C (NA4+VC)	NA4 (100 $\mu\text{g/ml}$ ) + VC (10 $\mu\text{g/ml}$ )

**Table 2:** Effect of the sodium arsenite and mitigating effects on viability of human lymphocytes measured by the trypan blue dye exclusion method.

[Values are mean $\pm$ SD]		
Groups	Treatment	Viability (%)
Group I	Control	95.63 $\pm$ 1.97
Group II	NA1	95.05 $\pm$ 1.34 <sup>abc</sup>
Group III	NA2	79.78 $\pm$ 2.03 <sup>***a</sup>
Group IV	NA3	77.00 $\pm$ 1.63 <sup>***b</sup>
Group V	NA4	56.34 $\pm$ 0.73 <sup>***cd</sup>
Group VI	VC	96.01 $\pm$ 0.41
Group VII	NA4 + VC	87.05 $\pm$ 4.88 <sup>***d</sup>

*P* Values: \*\*\* $P < 0.001$  compared to control group, <sup>a,b,c,d</sup> $< 0.001$  having similar superscripts in the respective columns are significantly different from each other.

### 3.2 Analysis of DNA damage by using comet assay

This study includes seven groups presented in Table 1. To study the dose dependent genotoxic effect induced by Sodium arsenite, four increasing doses were selected and comet assay was performed as endpoint of genotoxicity assessment. To study the beneficial effect of VC, the highest dose of sodium arsenite was given in VC pretreated lymphocytes as summarized in Table 3. A dose dependent increase in % tail DNA, Comet length and olive tail moment was observed, all of which were statistically significant when compared to untreated control lymphocytes. Increased percent DNA in comet tail ( $P < 0.01$ ;  $P < 0.001$ ), tail extent moment ( $P < 0.01$ ), olive tail moment ( $P < 0.01$ ;  $P < 0.001$ ) visibly indicated sodium arsenite induced DNA damage at the given treatment. This DNA damage by sodium arsenite escorted with the decreased percent DNA in comet tail ( $P < 0.05$ ) and olive tail moment ( $P < 0.01$ ) when supplemented with VC (10  $\mu\text{g/ml}$ ) in cultures. These findings visibly indicated protective action of VC against sodium arsenite induced DNA damage (Table 3).

**Table 3:** Comet parameter: % DNA in tail, tail extent moment and olive tail moment in human lymphocytes of control, sodium arsenite, vitamin C alone, and combined treatment of both sodium arsenite and vitamin c as measured by comet assay.

Treatment	[Values are mean $\pm$ SD]		
	% Tail DNA	Tail Extent Moment	Olive Tail Moment
Control	1.38 $\pm$ 0.30	0.03 $\pm$ 0.01	0.06 $\pm$ 0.02
NA1	2.73 $\pm$ 0.89 <sup>a</sup>	0.07 $\pm$ 0.03 <sup>b</sup>	0.10 $\pm$ 0.02 <sup>a</sup>
NA2	4.23 $\pm$ 1.55 <sup>**</sup>	0.16 $\pm$ 0.02	0.14 $\pm$ 0.05 <sup>**</sup>
NA3	5.13 $\pm$ 1.08 <sup>**</sup>	0.25 $\pm$ 0.05	0.19 $\pm$ 0.01 <sup>**</sup>
NA4	8.49 $\pm$ 1.17 <sup>***ab</sup>	0.32 $\pm$ 0.04 <sup>***b</sup>	0.27 $\pm$ 0.03 <sup>***a</sup>
VC	1.55 $\pm$ 0.05	0.02 $\pm$ 0.01	0.04 $\pm$ 0.01
NA4 + VC	6.28 $\pm$ 0.39 <sup>b</sup>	0.26 $\pm$ 0.06	0.25 $\pm$ 0.03 <sup>***a</sup>

P Values: \*\*\* $< 0.001$ ; \*\* $< 0.01$ ;  $P < 0.05$  compared to control group, <sup>b</sup> $< 0.05$ ; <sup>a</sup> $< 0.01$  values having similar superscripts in the respective columns are significantly different from each other.

## 4. DISCUSSION

The present study focuses on sodium arsenite induced genotoxicity and also the protective effect of VC on the lymphocytes *in vitro*. The enigma associated with arsenic is that it is point mutations and shows clastogenic effects in both animals and humans [19]. Epidemiological data gathered for more than a century have shown that arsenic and its species is a potent human carcinogen. Animal bioassays have failed to show any evidence of carcinogenesis. Arsenic is documented as a paradoxical human carcinogen that interacts indirectly with DNA [19].

In the current study, inorganic arsenic apparently caused dose-dependent increase in genotoxic indices and also cellular injury. VC is known to be an antioxidant and is able to scavenge free radicals. Earlier work suggests that VC and Vitamin E (VE) has proved evident as efficient drug against arsenic mediated toxicity [20]. Previous study confirmed the protective role of VE and VC on nickel induced liver damage, through alteration of liver marker enzymes and both vitamins have attenuated the toxic effects of nickel [21]. VC also helps to prevent atherosclerosis through the synthesis of collagen in the arterial wall and prevent

undesirable adhesion of leukocytes to the damaged artery [22]. Oral co-administration of rutin and ascorbic acid attenuates hypercholesterolemia-induced hepatic toxicity in female rats by decreasing liver enzymatic and lipid peroxidative markers as well as increasing the antioxidative cascade [23]. VE is an important lipid soluble antioxidant present in cells, as it is the major chain terminating antioxidant in biological membranes [24]. Previous *in vivo* and *in vitro* studies indicate that curcumin possesses antioxidant and anticarcinogenic properties [25, 26]. Earlier work indicates towards the protection of lymphocytes from toxic effects of arsenic by melatonin *in vitro* [27]. A recent study revealed that curcumin supplementation protects from *in vitro* genotoxic effects of arsenic in human lymphocytes [28].

Although how inorganic arsenic induces genetic injury is not fully explained, oxidative damage is likely involved in arsenic-induced DNA strand breaks, MN and even apoptosis in a variety of cell systems [29]. Ameliorative action of VC was observed against toxic effects of arsenic and Sodium fluoride, by reducing oxidative stress [30]. Arsenic induces reactive oxygen species (ROS) leading to oxidative stress and carcinogenesis [31]. It has been reported that arsenic exposure can result in increased cellular reactive oxygen species and oxidative DNA damage [32]. It has not been proven that inorganic arsenic reacts directly with DNA, but it has been suggested that arsenite induces oxidative damage causing single and double-strand breaks [33], although oxidative stress is only one of the possible mechanisms involved in arsenic genotoxicity.

Our study reveals the *in vitro* cytotoxicity of sodium arsenite in human lymphocytes. The Trypan blue dye exclusion test showed cytotoxic response in sodium arsenite treated human lymphocytes *in vitro* and indicating possible induction of mitochondrial damage by sodium arsenite. In the present study percent of cell viability decreased in all the doses of sodium arsenite treated lymphocytes. In the combined treatment of VC (10  $\mu\text{g/ml}$ ) and sodium arsenite (100  $\mu\text{g/ml}$ ) in the culture increased the cell viability and proving the positive effect of VC.

We found that VC at 10  $\mu\text{g/ml}$  significantly protected DNA of normal human lymphocytes against primary DNA damage induced by sodium arsenite at highest concentration (100  $\mu\text{g/ml}$ ). In this study, VC supplementation caused radical amelioration in the genotoxic effects brought about by sodium arsenite. This data thus signifies that VC has a definitive protective action against inorganic arsenic exerted genotoxicity (Table 3).

A recent study revealed the ability of arsenic and its metabolites to cause cytotoxic effects and primary DNA damage in human lymphocytes, when exposed *in vitro* for 24h, assessed by single cell gel electrophoresis as analyzed in the our study [34]. VC has antioxidant and free radical scavenging activities, which suggests that this vitamin may modulate oxidative DNA damage in mammalian cells [35]. Earlier finding shows that VC supplementation exhibits anti-hyperuricemic, anti-oxidation and nephroprotective activity in rat model of hyperuricemia-induced oxidative damage and renal injury [36]. It is observed that VC may

have hepatoprotective effect [37]. Arsenic contamination in ground water has become a major problem in many countries along with India. The problem is increasing day by day because there is no medicine that can prevent or cure arsenicosis. There is only measure to combat with arsenic related problems by providing arsenic-free water. Vitamins with anti-oxidant property viz., ascorbic acid found mainly in fresh vegetables can also be thought of as a dietary supplement. Our study has shown that sodium arsenite is a potent genotoxic agent capable of inducing DNA damage and also supported the beneficial effect of vitamin C in human lymphocytes *in vitro*.

## 5. CONCLUSION

Thus in view of present findings and previous works; we conclude that VC successfully ameliorates the *in vitro* genotoxic effects of sodium arsenite in human lymphocytes and clearly indicating its antioxidant property.

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## 7. REFERENCE

1. ATSDR. Toxicological Profile for Arsenic. US Department of Health and Human Services, Public Health Service, Atlanta, GA; 2005.
2. Tchounwou PB, Wilson BA, Ishaque A. Important considerations in the Development of public health advisories for arsenic and arsenic containing compounds in drinking water. *Rev on Environ. Health.* 1999; (14): 211.
3. Luchtrath H. Cirrhosis of the liver in chronic arsenical poisoning. *German Med Mon.* 1972; (2): 127.
4. Dipendra GM, R Haque, N Ghosh, Binoy KD, Santra A, Chakraborty D, Smith AH. Arsenic levels in drinking water and the prevalence of skin lesions in West Bengal, India. *Int J Epidemiol.* 1998; (27): 871.
5. Hayakawa T, Kobayashi Y, Cui X, Hirano S. A new metabolic pathway of arsenite: arsenic–glutathione complexes are substrates for human arsenic methyl transferase Cyt 19. *Arch Toxicol.* 2005; (79): 183.
6. Mandal BK, Ogra Y, Anzai K, Suzuki TK. Speciation of arsenic in biological samples. *Toxicol Appl Pharmacol.* 2004; (198): 307.
7. Tiwary H, Rao VM. Curcumin supplementation protects from genotoxic effects of arsenic and fluoride. *Food and Chemical Toxicology.* 2010; (48): 1234.
8. Smedley PL, Kinniburgh DG. A review of the source, behavior and distribution of arsenic in natural waters. *Appl Geochem.* 2002 (17): 517.
9. ATSDR. Department of Health and Human Services. Toxicological profile for arsenic; 2007.
10. Khuda-Bukhsh RA, Belon P, Boujedaini N, Pathak S, Bhattacharjee N, Bhattacharyya SS, Banerjee P. *Ecotoxicology and Environmental Safety.* 2009; (72): 639.
11. Monteiro HP, Stern A. Redox regulation of tyrosine phosphorylation-dependent signal transduction pathways. *Free Radic Biol Med.* 1996; (21): 323.
12. Nair SB, Jhala DD, Chinoy NJ. Mitigation of genotoxic effects of fluoride and arsenic by ascorbic acid in human lymphocyte cultures. *Fluoride.* 2004; (37): 249.
13. Singh S, Rana SV. Amelioration of arsenic toxicity by L-Ascorbic acid in laboratory rat. *J Environ Biol.* 2007; (28): 377.
14. Boyum A. Isolation of lymphocytes, granulocytes and macrophages. *Sc and J Immunol.* 1976; (5): 9.
15. Tennant JR. Evaluation of the trypan blue technique for determination of cell viability. *Transplantation.* 1964; (2): 685.
16. Singh NP, Mc Coy MT, Tice RR, Schneider EL. A simple technique for quantification of low levels of DNA damage in individual cells. *Exp Cell Res.* 1988; (175): 184.
17. Tice RR, Agurell E, Anderson D, Burlinson B, Hartmann A, Kobayashi H, Miyamae Y, Rojas E, Ryu JC, Sasaki YF. Single cell gel/comet assay: guidelines for *in vitro* and *in vivo* genetic toxicology testing. *Environ Mol Mutagen.* 2000; (35): 206.
18. Henderson L, Jones E, Brooks T, Chetelat A, Ciliutti P, Freemantle M, Howard CA, Mackay J, Phillips B, Riley S, Roberts C, Wotton AK, vande Waart EJ. Industrial genotoxicology group collaborative trial to investigate cell cycle parameters in human lymphocyte cytogenetic studies. *Mutagenesis.* 1997; (12): 163.
19. Basu A, Mahata J, Gupta S, Giri AK. Genetic toxicology of a paradoxical human carcinogen, arsenic-a review. *Mutat Res.* 2001; (488): 171.
20. Panneerselvam C, Samuel S, Muthuswamy A, Kadirvel R. Ascorbic acid and  $\alpha$ -tocopherol as potent modulators of apoptosis on arsenic induced toxicity in rats. *Toxicology Letters.* 2005; (156): 297.
21. Dahdouh F, Kechrid Z, Djebbar MR. Beneficial Effects of Vitamins (C + E) Supplementation against Nickel-induced Hepatotoxicity in Mice. *Advances in Bioresearch.* 2013; (4): 67.
22. Barrita JLS, Sánchez MSS. Antioxidant Role of Ascorbic Acid and His Protective Effects on Chronic Diseases. *Intech open science/open minds.* 2013; 449.
23. Aleisa AM, Abuhashish HM, Ahmed MM, Al-Rejaie SS, Alkhamees OA, Alroujaye AS. Ameliorative effects of rutin and ascorbic acid combination on hypercholesterolemia-induced hepatotoxicity in female rats. *African Journal of Pharmacy and Pharmacology.* 2013; (7): 280.
24. Burton GW, Joyce A, Ingold KU. Is vitamin E the only lipid-soluble, chain-breaking antioxidant in human blood plasma and erythrocyte membranes? *Arch Biochem Biophys.* 1983; (221): 281.
25. Naik RS, Majumdar AM, Ghaskadbi S. Protection of liver cells from ethanol cytotoxicity by curcumin in liver slice culture *in vitro.* *J Ethnopharmacol.* 2004; (95): 31.
26. Smerak P, Polivkova Z, Sestakova H, Stetina R, Barta I, Langova M, Turek B, Bartova J. Antimutagenic effect of curcumin and its effect on the immune response in mice. *Czech J Food Sci.* 2006; (24): 72.
27. Hemlata P, Rao H, Mandava V. Evaluation of *in vitro* anti-genotoxic potential of melatonin against arsenic and fluoride in human blood cultures. *Ecotoxicology and Environmental Safety.* 2010; (73): 1333.
28. Rao VM, Tiwari H. Curcumin supplementation protects from genotoxic effects of arsenic and fluoride. *Food and Chemical Toxicology.* 2010; (48): 1234.
29. Ho IH, Yih LH, Kao CY, Lee TC. Tin proto porphyrin potentiates arsenite-induced DNA strand breaks and kinetochore-negative micronuclei in human fibroblasts. *Mutat Res.* 2000; (452): 41.
30. Jhala DD, Chinoy NJ, Rao MV. Mitigating effects of some antidotes on fluoride and arsenic induced free radical toxicity in mice ovary. *Food Chem Toxicol.* 2008; (46): 1138.
31. Wang H, Xi S, Xu Y, Wang F, Zheng Y, Li B, Li X, Zheng Q, Sun G. Sodium arsenite induces cyclooxygenase-2 expression in human uroepithelial cells through MAPK pathway activation and reactive oxygen species induction. *Toxicol In Vitro.* 2013; (27): 1043.
32. Lai Y, Zhao W, Chen C, Wu M, Zhang Z. Role of DNA polymerase beta in the genotoxicity of arsenic. *Environ Mol Mutagen.* 2011; (52): 460.
33. Hei TK, Liu SX, Waldren C. Mutagenicity of arsenic in mammalian cells: role of reactive oxygen species. *Proc Natl Acad Sci USA.* 1998; (95): 8103.
34. Sordo M, Herrera LA, Ostrosky-Wegman P, Rojas E. Cytotoxic and genotoxic effects of As, MMA, and DMA on leukocytes and stimulated human lymphocytes. *Carcinog Mutagen.* 2001; (21): 249.
35. Odin AP. Vitamins as antimutagens: advantages and some possible mechanisms of antimutagenic action. *Mutat Res.* 1997; (386): 39.

36. Kensara OA. Protective effect of vitamin C supplementation on oxonate-induced hyperuricemia and renal injury in rats. *International journal of nutrition and metabolism*. 2013; (5): 61.
37. Adikwu E, Deo O. Hepatoprotective Effect of Vitamin C (Ascorbic Acid). *Pharmacology & Pharmacy*. 2013; 84.

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