

Antiproliferative activities of *Althaea ludwigii* L. extract on Michigan Cancer Foundation-7 breast cancer cell line

Zinah Essam Hameed Alshaya¹, Enas Jawad Kadhim¹, Hayder B. Sahib^{2*}

¹Department of Pharmacognosy, College of Pharmacy, University of Baghdad, Baghdad, Iraq, ²Department of Pharmacology, College of Pharmacy, Al-Nahrain University, Baghdad, Iraq

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ABSTRACT

Cancer has been considered one of these major community healthiness problems. *Althaea ludwigii* L. (family: Malvaceae) is an indigenous plant, widely distributed in Iraq. The objective of this study is to explore the antiproliferative action of *A. ludwigii* L. against breast cancer cell line Michigan Cancer Foundation-7 (MCF-7) and rate embryonic fibroblasts (REF) normal cell line and to identify the bioactive constituent. The aerial parts of *A. ludwigii* L. were grinded into fine powder then extracted with 85% methanol. Methanol extract was further partitioned with ethyl acetate. Ethyl acetate fraction was chosen to test against MCF-7 by (3-[4, 5-dimethylthiazol-2-yl]-2, 5 (MTT assay) the bioactive constituents were investigated by high performance thin layer chromatography high-performance thin-layer chromatography (HPTLC). The data showed that ethyl acetate fraction has significant antiproliferative activity against MCF-7 (IC₅₀ = 35.5 µg/ml) and REF (IC₅₀ = 1762.9 µg/ml), HPTLC showed the existence of rutin in ethyl acetate fraction. The antiproliferative activity may relate to the existence of rutin which had been approved as potent anticancer.

1. INTRODUCTION

For many years, medicinal plants had been the only method for the treatment of several ethnic groups; now, traditional medicinal plants are still used to treat or to act as prophylaxis against many diseases [1]. Many countries around the world use medicinal plants constituent as a common alternative for cancer prevention and treatment [2,3]. 60% of the anticancer drugs have been extracted from natural products; more than 3000 plants worldwide have been reported to possess anticancer properties [4]. Throughout history, plant extracts and their purified active components have been the backbone of cancer chemotherapeutics [5]. Cancer chemoprevention, by the use of natural, dietary, or synthetic agents that can reverse, suppress, or prevent carcinogenic progression, has become an appealing strategy to combat the dogma associated with increasing cases of cancers worldwide [6]. Various number of medicinal and poisonous plants grow in Iraq, which are mostly used for home remedies. Investigation of the active constituents of these plants might bring good credit for the drug industries; analysis of some wild drugs gave very satisfactory results [7]. Of these wildly grown and widely distributed plant species, Iraqi *Althaea ludwigii* L. (family: Malvaceae) was chosen for this study.

2. MATERIALS AND METHODS

2.1. Plant Material

A. ludwigii L. (Malvaceae) was harvested from Msayab city south of Baghdad. The plant was authenticated by the National Herbarium at Abu Ghraib. The plant was collected during the month of June (2017) and was rinsed with tap water, dried at room temperature in the shade, then pulverized by mechanical mills, the weight had been recorded.

2.2. Extraction

Shade-dried coarsely powdered aerial parts (200 g) were packed in a thimble of Soxhlet apparatus and extracted with 85% methanol until complete exhaustion. The alcoholic extract was filtered, and then, the solvent was evaporated under reduced pressure using the rotary evaporator at a 40°C. The extract was further partitioned with ethyl acetate. The fraction was dried over anhydrous sulfate, filtered, and evaporated to dryness. The fraction was weighted and assigned for further analysis [8].

2.3. Cell Cultures

Michigan Cancer Foundation-7 (MCF-7) cell line and rate embryonic fibroblasts (REF) cell line obtained from the Iraq biotech Cell Bank Unit and were maintained in RPMI-1640 supplemented with 10% fetal bovine, 100 units/mL penicillin, and 100 µg/mL streptomycin. Cells were incubated at 37°C and were [10].

*Corresponding Author
Hayder B. Sahib,
Department of Pharmacognosy,
College of Pharmacy, University of Baghdad,
Baghdad, Iraq. H.P.: 00964-7812353838.
Email: haider_bahaa@yahoo.com

2.3.1. Cell viability assay

The cell viability was evaluated by thiazolyl blue tetrazolium bromide (MTT) assay. Cells were seeded in 96-well plates at concentration of 1×10^4 /well, passage 8 and incubated at 37°C for 24 h. Then, MEOS was added by dissolving the fraction dimethyl sulfoxide (DMSO) and diluted the mixture with the medium to make the final DMSO concentration of 1% MEOS was incubated with the cells for 72 h, and then, 20 μ l from 5 mg/mL of MTT was added on each well and incubated for 5 h, at 37°C and 5% CO₂. Then, medium was aspirated and 200 μ l from DMSO added to dissolve the formazan crystal that is formed. The spectrophotometric absorbance of the samples had been measured by the microplate readers, with a measurement wavelength of 570 nm and reference wavelength of 650 nm. DMSO at 1% was used as negative control and the results were presented as mean percent of inhibition to the negative control \pm SD. Each experiment was performed in quadruplicate different concentrations of the ethyl acetate fraction (6.25, 12.5, 25, 50, and 100 μ g/ml) used [9].

In a study by Sahib *et al.* [11], the inhibitory effect was calculated according to the following equation:

$$\text{Cell growth inhibition (\%)} = (1 - (A_0/A)) \times 100$$

Where,

A₀ = the absorbance of the samples

A = the absorbance of the negative control

2.4. High-performance Thin-layer Chromatography (HPTLC)

Flavonoids and phenolic acids in the fraction were identified using HPTLC.

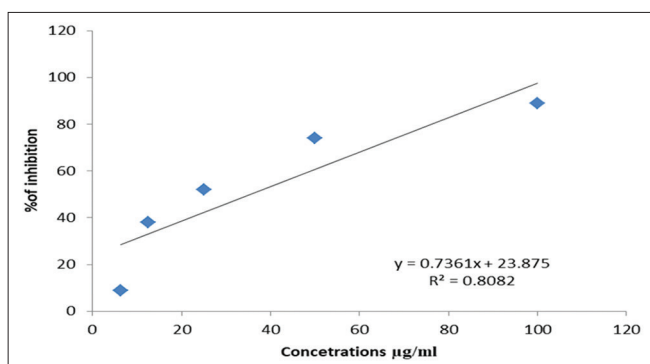


Figure 1: Antiproliferative activity of *Althaea ludwigii* against Michigan Cancer Foundation-7

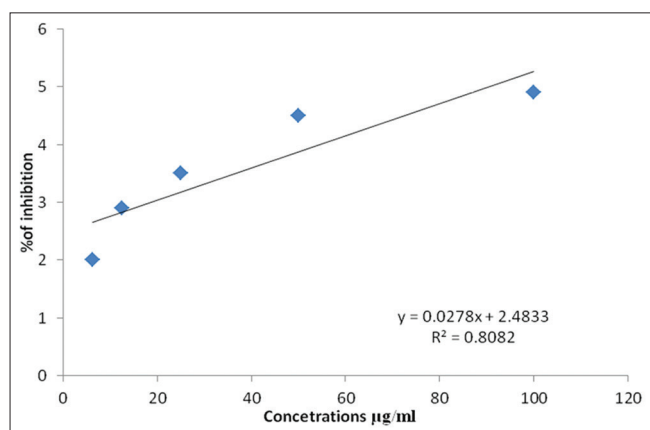


Figure 2: Antiproliferative activity of *Althaea ludwigii* against rate embryonic fibroblasts

2.4.1. Chromatographic conditions

Standard and sample solutions (rutin, quercetin, myricetin, kaempferol, silymarin, and ethyl acetate F.) (5.0 μ L of each concentration 1 mg/mL) were applied to the plates 5 μ L each, using Camag Linomat V automatic sample applicator in the form of band (band length: 8 mm) using microsyringe. The plates were saturated for 30 min in a twin trough glass chamber (for 10 \times 10 cm) with the mobile phase of ethyl acetate:formic acid:acetic acid:water (100:11:11:27) subsequent to the development [12,13]. The plates were air dried and scanning (slit dimensions: 5 \times 0.45) was performed at 254 nm and 366 nm using Camag TLC Scanner III.

3. RESULTS AND DISCUSSION

The results in Figure 1 showed that ethyl acetate fraction of *A. ludwigii* L. had significant antiproliferative activity against MCF-7 hormonal-dependent cancer cell line, the IC₅₀ was 35.5 μ g/mL. However, the results in Figure 2 showed that the IC₅₀ was 1709.23 μ g/ml on REF cell line which is normal cell line, the data may indicate that *A. ludwigii* L. may target the cancer cell only rather than cancer cell, this promising results may solve a big problem facing clinicians when they prescribing chemotherapy, as the chemotherapy killing both normal and cancer cells, it was important either to find an alternatives or to find an adjuvants agents which may help in reducing the chemotherapy dose and to act by mechanism(s) rather than the chemotherapy mechanisms [14]. The potential antiproliferative activity of *A. ludwigii* L. ethyl acetate fraction made sense to the researches to search for the bioactive constituents which may be responsible on the antiproliferative activity. Ethyl acetate fraction of the *A. ludwigii* L.

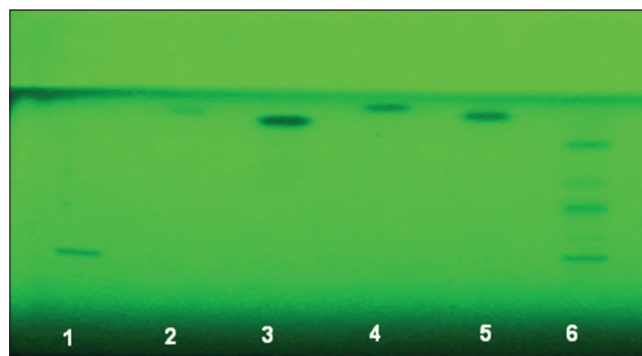


Figure 3: Chromatogram obtained from standards (1: Rutin, 2: Quercetin, 3: Myricetin, 4: Kaempferol, 5: Silymarin, and sample 6: Ethyl acetate F.). Visualization was under UV light of wavelength 254 nm

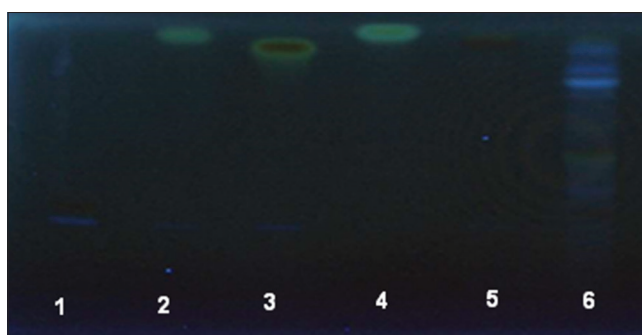


Figure 4: Chromatogram obtained from standards (1: Rutin, 2: Quercetin, 3: Myricetin, 4: Kaempferol, 5: Silymarin, and sample 6: Ethyl acetate F.). Visualization was under UV light of wavelength 366 nm

extract was nominated to be investigated for the bioactive constituents using HPTLC; the results showed by Figures 3 and 4 that rutin has been identified in ethyl acetate fraction. A previous study done by Sahib *et al.*, Chen *et al.* [15,16] showed that rutin significantly inhibited the proliferation of LAN-5 cells line. Rutin had been approved that it induced G2/M arrest in the cell cycle progression and induced cell apoptosis. Moreover, rutin could decrease BCL2 expression and BCL2/BAX ratio as well. In the meantime, the MYCN mRNA level and the secretion of tumor necrosis factor- α were inhibited [15]. In conclusion, these results suggest that rutin produces clear antiproliferative effects through encouraged G2/M arrest in the cell cycle development and induced cell apoptosis moreover, regulating the expression of the gene linked to apoptosis [15].

4. CONCLUSION

A. ludwigii L. extract may be has promising anti-breast cancer activity, with little effect on normal cells. The activity may related to the flavonoid exist in the extract.

5. ACKNOWLEDGMENTS

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